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(57) Abstract

The present invention relates to polynucleotide and polypeptide molecules for mammalian alpha helix-12 (Zalpha12). The polypeptides, and polynucleotides encoding them, are hormonal and may be used to regulate the functioning of the immune system. The present invention also includes antibodies to the Zalpha12 polypeptides. Antagonists may be used to treat inflammation and inflammation-related diseases.

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MAMMALIAN ALPHA-HELICAL PROTEIN - 12

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BACKGROUND OF THE INVENTION

Inflammation normally is a localized, protective response to trauma or microbial invasion that destroys,

10 dilutes, or walls-off the injurious agent and the injured tissue. It is characterized in the acute form by the classic signs of pain, heat, redness, swelling, and loss of function. Microscopically it involves a complex series of events, including dilation of arterioles, capillaries, and venules, with increased permeability and blood flow, exudation of fluids, including plasma proteins, and leukocyte migration into the area of inflammation.

Diseases characterized by inflammation are significant causes of morbidity and mortality in humans. 20 Commonly, inflammation occurs as a defensive response to invasion of the host by foreign, particularly microbial, material. Responses to mechanical trauma, toxins, and neoplasia also may results in inflammatory reactions. The accumulation and subsequent activation of leukocytes are 25 central events in the pathogenesis of most forms of inflammation. Deficiencies of inflammation compromise the host. Excessive inflammation caused by abnormal recognition of host tissue as foreign or prolongation of the inflammatory process may lead to inflammatory diseases. 30 as diverse as diabetes, arteriosclerosis, cataracts, reperfusion injury, and cancer, to post-infectious syndromes such as in infectious meningitis, rheumatic fever, and to rheumatic diseases such as systemic lupus erythematosus and rheumatoid arthritis. The centrality of 35 the inflammatory response in these varied disease processes makes its regulation a major element in the

prevention control or cure of human disease. Thus, there

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is a need to discover cytokines which contribute to inflammation and inflammatory related diseases so that antagonists such as antibodies can be administered to down-regulate the cytokine so as to ameliorate the inflammation.

SUMMARY OF THE INVENTION

The present invention addresses this need by providing novel polypeptides and related compositions and methods and their antagonists. Within one aspect, the present invention provides an isolated polynucleotide encoding a mammalian cytokine termed Zalpha12. The data show that the cytokine is involved in the inflammation response. Thus, antagonists of Zalpha12 can be used to lessen inflammation especially inflammation associated with coronary heart disease, arteriosclerosis, Crohn's disease and inflammatory bowel disease.

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Three variants have been discovered as shown in SEQ ID NOs:1, 2, 3, 4, 5 and 6. Each Zalpha12 polypeptide has a signal sequence which extends from amino acid residue 1, a methionine to and including amino acid 25 residue 34, a serine of SEQ ID NOs:2, 4 and 6. Thus, the Zalpha12 polypeptide represented by SEQ ID NO:2 has a mature sequence extending from amino acid residue 35, an alanine, to and including amino acid residue 202, an asparagine, also represented by SEQ ID NO:8. The Zalpha12 30 polypeptide represented by SEQ ID NO:4 has a mature sequence extending from amino acid residue 35, an alanine to and including amino acid residue 288, an asparagine also represented by SEQ ID NO:9. The Zalpha12 polypeptide represented by SEQ ID NO:6 has a mature sequence extending 35 from amino acid residue 35, an alanine to and including amino acid residue 158, an asparagine, also represented by SEQ ID NO:10.

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Within a second aspect of the invention there is provided an expression vector comprising (a) a transcription promoter; (b) a DNA segment encoding Zalpha12 polypeptide, and (c) a transcription terminator, wherein the promoter, DNA segment, and terminator are operably linked.

Within a third aspect of the invention there is provided a cultured eukaryotic or prokaryotic cell into which has been introduced an expression vector as disclosed above, wherein said cell expresses a protein polypeptide encoded by the DNA segment.

Within a further aspect of the invention there is provided a chimeric polypeptide consisting essentially of a first portion and a second portion joined by a peptide bond. The first portion of the chimeric polypeptide consists essentially of (a) a Zalpha12 polypeptide (b) allelic variants of Zalpha12; and (c) protein polypeptides that are at least 80% identical to (a) or (b). The second portion of the chimeric polypeptide consists essentially of another polypeptide such as an affinity tag. Within one embodiment the affinity tag is an immunoglobulin F_C polypeptide. The invention also provides expression vectors encoding the chimeric polypeptides and host cells transfected to produce the chimeric polypeptides.

Within an additional aspect of the invention there is provided an antibody that specifically binds to a Zalpha12 polypeptide as disclosed above, and also an anti-idiotypic antibody which neutralizes the antibody to a Zalpha12 polypeptide.

An additional embodiment of the present invention relates to a peptide or polypeptide which has the amino acid sequence of an epitope-bearing portion of a Zalpha12 polypeptide having an amino acid sequence described above. Peptides or polypeptides having the amino acid sequence of an epitope-bearing portion of a Zalpha12

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polypeptide of the present invention include portions of such polypeptides with at least nine, preferably at least 15 and more preferably at least 30 to 50 amino acids, although epitope-bearing polypeptides of any length up to and including the entire amino acid sequence of a polypeptide of the present invention described above are also included in the present invention. Examples of such epitope binding regions are SEQ ID NOs: 18, 19, 20, 25, 26, 27, 28, 29, 30, 35, 36, 37, 38, 39, 40, 41, 42, 43 and 44. Also claimed are any of these polypeptides that are fused to another polypeptide or carrier molecule.

These and other aspects of the invention will become evident upon reference to the following detailed description of the invention.

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DETAILED DESCRIPTION OF THE INVENTION

The teachings of all the references cited herein are incorporated in their entirety herein by reference.

Prior to setting forth the invention in detail, it may be helpful to the understanding thereof to define the following terms:

The term "affinity tag" is used herein to denote a polypeptide segment that can be attached to a second polypeptide to provide for purification or detection of the second polypeptide or provide sites for attachment of the second polypeptide to a substrate. In principal, any peptide or protein for which an antibody or other specific binding agent is available can be used as an affinity tag.

30 Affinity tags include a poly-histidine tract, protein A, Nilsson et al., EMBO J. 4:1075 (1985); Nilsson et al., Methods Enzymol. 198:3 (1991), glutathione S transferase, Smith and Johnson, Gene 67:31 (1988), Glu-Glu affinity tag, Grussenmeyer et al., Proc. Natl. Acad. Sci. USA

35 82:7952-4 (1985), substance P, Flag™ peptide, Hopp et al., Biotechnology 6:1204-1210 (1988), streptavidin binding

peptide, or other antigenic epitope or binding domain. See, in general, Ford et al., Protein Expression and Purification 2: 95-107 (1991). DNAs encoding affinity tags are available from commercial suppliers (e.g., 5 Pharmacia Biotech, Piscataway, NJ).

The term "allelic variant" is used herein to denote any of two or more alternative forms of a gene occupying the same chromosomal locus. Allelic variation arises naturally through mutation, and may result in phenotypic polymorphism within populations. Gene mutations can be silent (no change in the encoded polypeptide) or may encode polypeptides having altered amino acid sequence. The term allelic variant is also used herein to denote a protein encoded by an allelic variant of a gene.

The terms "amino-terminal" and "carboxylterminal" are used herein to denote positions within
polypeptides. Where the context allows, these terms are
used with reference to a particular sequence or portion of
a polypeptide to denote proximity or relative position.
For example, a certain sequence positioned carboxylterminal to a reference sequence within a polypeptide is
located proximal to the carboxyl terminus of the reference
sequence, but is not necessarily at the carboxyl terminus
of the complete polypeptide.

The term "complement/anti-complement pair"

denotes non-identical moieties that form a non-covalently
associated, stable pair under appropriate conditions. For
instance, biotin and avidin (or streptavidin) are

prototypical members of a complement/anti-complement pair.
Other exemplary complement/anti-complement pairs include
receptor/ligand pairs, antibody/antigen (or hapten or
epitope) pairs, sense/antisense polynucleotide pairs, and
the like. Where subsequent dissociation of the
complement/anti-complement pair is desirable, the

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complement/anti-complement pair preferably has a binding affinity of $<10^9 M^{-1}$.

The term "complements of a polynucleotide molecule" is a polynucleotide molecule having a 5 complementary base sequence and reverse orientation as compared to a reference sequence. For example, the sequence 5' ATGCACGGG 3' is complementary to 5' CCCGTGCAT 31.

The term "contig" denotes a polynucleotide that 10 has a contiguous stretch of identical or complementary sequence to another polynucleotide. Contiguous sequences are said to "overlap" a given stretch of polynucleotide sequence either in their entirety or along a partial stretch of the polynucleotide. For example,

15 representative contigs to the polynucleotide sequence 5'-ATGGCTTAGCTT-3' are 5'-TAGCTTgagtct-3' and 3'gtcgacTACCGA-5'.

The term "degenerate nucleotide sequence" denotes a sequence of nucleotides that includes one or 20 more degenerate codons (as compared to a reference polynucleotide molecule that encodes a polypeptide). Degenerate codons contain different triplets of nucleotides, but encode the same amino acid residue (i.e., GAU and GAC triplets each encode Asp).

The term "expression vector" is used to denote a DNA molecule, linear or circular, that comprises a segment encoding a polypeptide of interest operably linked to additional segments that provide for its transcription. Such additional segments include promoter and terminator 30 sequences, and may also include one or more origins of replication, one or more selectable markers, an enhancer, a polyadenylation signal, etc. Expression vectors are generally derived from plasmid or viral DNA, or may contain elements of both.

35 The term "isolated", when applied to a polynucleotide, denotes that the polynucleotide has been removed from its natural genetic milieu and is thus free

of other extraneous or unwanted coding sequences, and is in a form suitable for use within genetically engineered protein production systems. Such isolated molecules are those that are separated from their natural environment and include cDNA and genomic clones. Isolated DNA molecules of the present invention are free of other genes with which they are ordinarily associated, but may include naturally occurring 5' and 3' untranslated regions such as promoters and terminators. The identification of associated regions will be evident to one of ordinary skill in the art (see for example, Dynan and Tijan, Nature 316:774-78 (1985).

An "isolated" polypeptide or protein is a

15 polypeptide or protein that is found in a condition other
than its native environment, such as apart from blood and
animal tissue. In a preferred form, the isolated
polypeptide is substantially free of other polypeptides,
particularly other polypeptides of animal origin. It is

20 preferred to provide the polypeptides in a highly purified
form, i.e. greater than 95% pure, more preferably greater
than 99% pure. When used in this context, the term
"isolated" does not exclude the presence of the same
polypeptide in alternative physical forms, such as dimers

25 or alternatively glycosylated or derivatized forms.

The term "operably linked", when referring to DNA segments, indicates that the segments are arranged so that they function in concert for their intended purposes, e.g., transcription initiates in the promoter and proceeds through the coding segment to the terminator.

The term "ortholog" denotes a polypeptide or protein obtained from one species that is the functional counterpart of a polypeptide or protein from a different species. Sequence differences among orthologs are the result of speciation.

"Paralogs" are distinct but structurally related proteins made by an organism. Paralogs are believed to arise through gene duplication. For example, a-globin, b-globin, and myoglobin are paralogs of each other.

- A "polynucleotide" is a single- or doublestranded polymer of deoxyribonucleotide or ribonucleotide bases read from the 5' to the 3' end. Polynucleotides include RNA and DNA, and may be isolated from natural sources, synthesized in vitro, or prepared from a
- 10 combination of natural and synthetic molecules. Sizes of polynucleotides are expressed as base pairs (abbreviated "bp"), nucleotides ("nt"), or kilobases ("kb"). Where the context allows, the latter two terms may describe polynucleotides that are single-stranded or double-
- stranded. When the term is applied to double-stranded molecules it is used to denote overall length and will be understood to be equivalent to the term "base pairs". It will be recognized by those skilled in the art that the two strands of a double-stranded polynucleotide may differ
- slightly in length and that the ends thereof may be staggered as a result of enzymatic cleavage; thus all nucleotides within a double-stranded polynucleotide molecule may not be paired. Such unpaired ends will in general not exceed 20 nt in length.
- A "polypeptide" is a polymer of amino acid residues joined by peptide bonds, whether produced naturally or synthetically. Polypeptides of less than about 10 amino acid residues are commonly referred to as "peptides".
- The term "promoter" is used herein for its artrecognized meaning to denote a portion of a gene
 containing DNA sequences that provide for the binding of
 RNA polymerase and initiation of transcription. Promoter
 sequences are commonly, but not always, found in the 5'
 non-coding regions of genes.

A "protein" is a macromolecule comprising one or more polypeptide chains. A protein may also comprise non-

peptidic components, such as carbohydrate groups.

Carbohydrates and other non-peptidic substituents may be added to a protein by the cell in which the protein is produced, and will vary with the type of cell. Proteins are defined herein in terms of their amino acid backbone structures; substituents such as carbohydrate groups are generally not specified, but may be present nonetheless.

The term "receptor" denotes a cell-associated protein that binds to a bioactive molecule (i.e., a ligand) and mediates the effect of the ligand on the cell. Membrane-bound receptors are characterized by a multidomain structure comprising an extracellular ligandbinding domain and an intracellular effector domain that is typically involved in signal transduction. Binding of ligand to receptor results in a conformational change in the receptor that causes an interaction between the effector domain and other molecule(s) in the cell. interaction in turn leads to an alteration in the metabolism of the cell. Metabolic events that are linked 20 to receptor-ligand interactions include gene transcription, phosphorylation, dephosphorylation, increases in cyclic AMP production, mobilization of cellular calcium, mobilization of membrane lipids, cell adhesion, hydrolysis of inositol lipids and hydrolysis of phospholipids. In general, receptors can be membrane bound, cytosolic or nuclear; monomeric (e.g., thyroid stimulating.hormone receptor, beta-adrenergic receptor) or multimeric (e.g., PDGF receptor, growth hormone receptor, IL-3 receptor, GM-CSF receptor, G-CSF receptor, erythropoietin receptor and IL-6 receptor). 30

The term "secretory signal sequence" denotes a
DNA sequence that encodes a polypeptide (a "secretory
peptide") that, as a component of a larger polypeptide,
directs the larger polypeptide through a secretory pathway
of a cell in which it is synthesized. The larger
polypeptide is commonly cleaved to remove the secretory
peptide during transit through the secretory pathway.

The term "splice variant" is used herein to denote alternative forms of RNA transcribed from a gene. Splice variation arises naturally through use of alternative splicing sites within a transcribed RNA molecule, or less commonly between separately transcribed RNA molecules, and may result in several mRNAs transcribed from the same gene. Splice variants may encode polypeptides having altered amino acid sequence. The term splice variant is also used herein to denote a protein encoded by a splice variant of an mRNA transcribed from a gene.

Molecular weights and lengths of polymers determined by imprecise analytical methods (e.g., gel electrophoresis) will be understood to be approximate values. When such a value is expressed as "about" X or "approximately" X, the stated value of X will be understood to be accurate to ±10%.

Introduction

20 The Zalpha12 protein is an alpha helical protein. SEQ ID NO:2 has four alpha helices, A, B, C and D. Helix A of SEQ ID NO:2 extends from amino acid residue 56, a leucine, to and including amino acid residue 70, an isoleucine also defined by SEQ ID NO:21. Helix B of SEQ ID NO:2 extends 25 from amino acid residue 96, a leucine to and including amino acid residue 110, a tyrosine, also defined by SEQ ID NO:22. Helix C of SEQ ID NO:2 extends from amino acid residue 130, a leucine, to and including amino acid residue 144, a leucine, also defined by SEQ ID NO:23. 30 Helix D of SEQ ID NO: 2 extends from amino acid residue 162, a methionine, to and including amino acid residue 176, a serine, also defined by SEQ ID NO:24. SEQ ID NO:25 contains helices A + B; SEQ ID NO:26 contains helices A + B + C; SEQ ID NO:27 contains helices A + B + C + D; SEQ ID 35 NO:28 contains helices B + C + D; SEQ ID NO:29 contains helices B + C; and SEQ ID NO:30 contains helices C + D of SEQ ID NO:2.

The polypeptide of SEQ ID NO:4 also contains four helices A, B, C and D. Helix A of SEQ ID NO:4 extends from amino acid residue 45, a histidine, to and including amino acid residue 59, a leucine, also defined by SEQ ID NO:31. 5 Helix B of SEQ ID NO:4 extends from amino acid residue 116, a valine, to and including amino acid residue 130, a lysine, also defined by SEQ ID NO:32. Helix C of SEQ ID NO:4 extends from amino acid residue 142, a leucine, to and including amino acid residue 156, an isoleucine, also 10 defined by SEQ ID NO:33. Helix D of SEQ ID NO:4 extends from amino acid residue 182, a leucine, to and including amino acid residue 196, a tyrosine, also defined by SEQ ID NO:34. SEQ ID NO:35 contains helices A + B of SEQ ID NO:4. SEQ ID NO:36 contains helices A + B + C of SEQ ID NO:4. 15 SEQ ID NO:37 contains helices A + B + C + D of SEQ ID NO:4. SEQ ID NO: 38 contains helices B + C + D of SEQ ID NO:4. SEQ ID NO:39 contains helices B + C of SEQ ID NO:4.

Expression of the Zalpha12 gene is seen in a number

of different tissues including the spleen, thymus, testis, small intestines, colon, peripheral blood lymphocytes (PBL), stomach, trachea, T-cells including CD4+ and CD8+ cells, and bone marrow. This pattern of expression suggests that zalpha12 may play a general role in development and exert important regulatory control of testicular differentiation, of the hypothalamic-pituitary-gonadal axis, and of gonadal steroidogenesis and spermatogenesis.

SEQ ID NO:40 contains helices C + D of SEQ ID NO:4.

Development of testicular hormone production can be divided into early and late steps, with the latter dependent on the activation of functionally-determined Leydig cell precursors by luteinizing hormone (LH). However, the factors that control the early steps in this process remain unknown, Huhtaniemi, Reprod. Fertil. Dev.

35 7: 1025-1035 (1995) suggesting that zalpha12 might be

responsible for activation of a non-steroidogenic, non-LH responsive precursor cell.

Once Leydig cell differentiation has occurred, production of steroid hormones in the testis is dependent 5 on the secretion of the gonadotrophins, LH and folliclestimulating hormone (FSH), by the pituitary. stimulates production of testosterone by the Leydig cells, whereas spermatogenesis depends on both FSH and high intratesticular testosterone concentrations. LH and FSH 10 secretion is in turn under control of gonadotrophin releasing hormone (GnRH) produced in the hypothalamus, Kaufman, The neuro endocrine regulation of male reproduction. in: Male Infertility. Clinical Investigation, Cause Evaluation and Treatment., FH 15 Comhaire, ed., pp 29-54 (Chapman and Hall, London, 1996). Since testicular products have been shown to control LH and FSH production, this suggests that zalpha12 might regulate hormone production by the hypothalamus.

It is well known that steroidogenesis and spermatogenesis take place within two different cellular compartments of the testes, with Leydig and Sertoli cells responsible for the former and latter, respectively, Saez , Endocrin. Rev. 15: 574-626 (1994). The activity of each of these cell types appears to be regulated by the 25 secretory products of the other. Sertoli cell derived tumor necrosis factor-a, fibroblast growth factor, interleukin-1, transforming growth factor-B, epidermal growth factor/transforming growth factor-a, activin, inhibin, insulin-like growth factor-1, platelet derived 30 growth factor, endothelin, and ariginine-vasopressin have all been shown to regulate Leydig cell function, Saez , Endocrin. Rev. 15: 574-626 (1994). Thus, zalpha12 might control or modulate the activities of one or more of these genes.

In men, aging is associated with a progressive decline in testicular function. These changes are

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manifest clinically by decreased virility, vigor, and libido that point towards a relative testicular deficiency, Vermeulen, Ann. Med. 25:531-534 (1993); Pugeat et al., Horm. Res. 43: 104-110 (1995). Hormone

5 replacement therapy in elderly men is not currently recommended which suggests that a new therapy for the male climacterium would be very valuable.

POLYNUCLEOTIDES:

The present invention also provides polynucleotide molecules, including DNA and RNA molecules, that encode the Zalpha12 polypeptides disclosed herein. Those skilled in the art will readily recognize that, in view of the degeneracy of the genetic code, considerable sequence variation is possible among these polynucleotide molecules.

Polynucleotides, generally a cDNA sequence, of the present invention encode the described polypeptides

20 herein. A cDNA sequence which encodes a polypeptide of the present invention is comprised of a series of codons, each amino acid residue of the polypeptide being encoded by a codon and each codon being comprised of three nucleotides. The amino acid residues are encoded by their respective codons as follows.

Alanine (Ala) is encoded by GCA, GCC, GCG or GCT;

Cysteine (Cys) is encoded by TGC or TGT;

Aspartic acid (Asp) is encoded by GAC or GAT;

Glutamic acid (Glu) is encoded by GAA or GAG;

Phenylalanine (Phe) is encoded by TTC or TTT;

Glycine (Gly) is encoded by GGA, GGC, GGG or

GGT;

Histidine (His) is encoded by CAC or CAT;
Isoleucine (Ile) is encoded by ATA, ATC or ATT;

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CCT;

ACT;

Lysine (Lys) is encoded by AAA, or AAG;

Leucine (Leu) is encoded by TTA, TTG, CTA, CTC,

CTG or CTT;

Methionine (Met) is encoded by ATG;
Asparagine (Asn) is encoded by AAC or AAT;
Proline (Pro) is encoded by CCA, CCC, CCG or

Glutamine (Gln) is encoded by CAA or CAG;
Arginine (Arg) is encoded by AGA, AGG, CGA, CGC,
10 CGG or CGT;

Serine (Ser) is encoded by AGC, AGT, TCA, TCC, TCG or TCT;

Threonine (Thr) is encoded by ACA, ACC, ACG or

Valine (Val) is encoded by GTA, GTC, GTG or GTT;
Tryptophan (Trp) is encoded by TGG; and
Tyrosine (Tyr) is encoded by TAC or TAT.

It is to be recognized that according to the
present invention, when a polynucleotide is claimed as
described herein, it is understood that what is claimed
are both the sense strand, the anti-sense strand, and the
DNA as double-stranded having both the sense and antisense strand annealed together by their respective

- 25 hydrogen bonds. Also claimed is the messenger RNA (mRNA) which encodes the polypeptides of the president invention, and which mRNA is encoded by the cDNA described herein.

 Messenger RNA (mRNA) will encode a polypeptide using the same codons as those defined herein, with the exception
- 30 that each thymine nucleotide (T) is replaced by a uracil nucleotide (U).

One of ordinary skill in the art will also appreciate that different species can exhibit "preferential codon usage." In general, see, Grantham, et al., Nuc. Acids Res. 8:1893-1912 (1980); Haas, et al. Curr. Biol. 6:315-324 (1996); Wain-Hobson, et al., Gene

13:355-364 (1981); Grosjean and Fiers, Gene 18:199-209 (1982); Holm, Nuc. Acids Res. 14:3075-3087 (1986); Ikemura, J. Mol. Biol. 158:573-597 (1982). As used herein, the term "preferential codon usage" or 5 "preferential codons" is a term of art referring to protein translation codons that are most frequently used in cells of a certain species, thus favoring one or a few representatives of the possible codons encoding each amino acid (See Table 2). For example, the amino acid Threonine 10 (Thr) may be encoded by ACA, ACC, ACG, or ACT, but in mammalian cells ACC is the most commonly used codon; in other species, for example, insect cells, yeast, viruses or bacteria, different Thr codons may be preferential. Preferential codons for a particular species can be 15 introduced into the polynucleotides of the present invention by a variety of methods known in the art. Introduction of preferential codon sequences into recombinant DNA can, for example, enhance production of the protein by making protein translation more efficient 20 within a particular cell type or species. Sequences containing preferential codons can be tested and optimized for expression in various species, and tested for functionality as disclosed herein.

Within preferred embodiments of the invention

25 the isolated polynucleotides will hybridize to similar sized regions of SEQ ID NOs:1, 3 or 5 or a sequence complementary thereto, under stringent conditions. In general, stringent conditions are selected to be about 5°C lower than the thermal melting point (T_m) for the specific sequence at a defined ionic strength and pH. The T_m is the temperature (under defined ionic strength and pH) at which 50% of the target sequence hybridizes to a perfectly matched probe. Typical stringent conditions are those in which the salt concentration is up to about 0.03 M at pH 7

35 and the temperature is at least about 60°C.

As previously noted, the isolated polynucleotides of the present invention include DNA and Methods for preparing DNA and RNA are well known in In general, RNA is isolated from a tissue or 5 cell that produces large amounts of Zalpha12 RNA. tissues and cells are identified by Northern blotting, Thomas, Proc. Natl. Acad. Sci. USA 77:5201 (1980) and are discussed below. Total RNA can be prepared using quanidine HCl extraction followed by isolation by centrifugation in 10 a CsCl gradient, Chirgwin et al., Biochemistry 18:52-94 (1979). Poly (A) $^+$ RNA is prepared from total RNA using the method of Aviv and Leder, Proc. Natl. Acad. Sci. USA 69:1408-1412 (1972). Complementary DNA (cDNA) is prepared from poly(A) + RNA using known methods. In the 15 alternative, genomic DNA can be isolated. Polynucleotides · encoding Zalpha12 polypeptides are then identified and isolated by, for example, hybridization or PCR.

A full-length clone encoding Zalpha12 can be obtained by conventional cloning procedures.

Complementary DNA (cDNA) clones are preferred, although for some applications (e.g., expression in transgenic animals) it may be preferable to use a genomic clone, or to modify a cDNA clone to include at least one genomic intron. Methods for preparing cDNA and genomic clones are well known and within the level of ordinary skill in the art, and include the use of the sequence disclosed herein, or parts thereof, for probing or priming a library. Expression libraries can be probed with antibodies to Zalpha12, receptor fragments, or other specific binding partners.

The polynucleotides of the present invention can also be synthesized using DNA synthesizers. Currently the method of choice is the phosphoramidite method. If chemically synthesized double stranded DNA is required for an application such as the synthesis of a gene or a gene fragment, then each complementary strand is made

separately. The production of short genes (60 to 80 bp) is technically straightforward and can be accomplished by synthesizing the complementary strands and then annealing them. For the production of longer genes (>300 bp),

5 however, special strategies must be invoked, because the coupling efficiency of each cycle during chemical DNA synthesis is seldom 100%. To overcome this problem, synthetic genes (double-stranded) are assembled in modular form from single-stranded fragments that are from 20 to 100 nucleotides in length.

See Glick and Pasternak, Molecular

Biotechnology, Principles & Applications of Recombinant

DNA, (ASM Press, Washington, D.C. 1994); Itakura et al.,

Annu. Rev. Biochem. 53: 323-356 (1984) and Climie et al.,

Proc. Natl. Acad. Sci. USA 87:633-637 (1990).

The present invention further provides counterpart polypeptides and polynucleotides from other species (orthologs). These species include, but are not limited to mammalian, avian, amphibian, reptile, fish, 20 insect and other vertebrate and invertebrate species. particular interest are Zalpha12 polypeptides from other mammalian species, including murine, porcine, ovine, bovine, canine, feline, equine, and other primate polypeptides. Orthologs of human Zalpha12 can be cloned 25 using information and compositions provided by the present invention in combination with conventional cloning techniques. For example, a cDNA can be cloned using mRNA obtained from a tissue or cell type that expresses Zalpha12 as disclosed herein. Suitable sources of mRNA 30 can be identified by probing Northern blots with probes designed from the sequences disclosed herein. A library is then prepared from mRNA of a positive tissue or cell A Zalpha12-encoding cDNA can then be isolated by a variety of methods, such as by probing with a complete or 35 partial human cDNA or with one or more sets of degenerate probes based on the disclosed sequences. A cDNA can also

be cloned using the polymerase chain reaction, or PCR (Mullis, U.S. Patent No. 4,683,202), using primers designed from the representative human Zalpha12 sequence disclosed herein. Within an additional method, the cDNA library can be used to transform or transfect host cells, and expression of the cDNA of interest can be detected with an antibody to Zalpha12 polypeptide. Similar techniques can also be applied to the isolation of genomic clones.

10 Those skilled in the art will recognize that the sequence disclosed in SEQ ID NOs:1, 3 or 5 represent specific alleles of human Zalpha12 and that allelic variation and alternative splicing are expected to occur. Allelic variants of this sequence can be cloned by probing 15 cDNA or genomic libraries from different individuals according to standard procedures. Allelic variants of the DNA sequence shown in SEQ ID NO:1, including those containing silent mutations and those in which mutations result in amino acid sequence changes, are within the 20 scope of the present invention, as are proteins which are allelic variants of SEQ ID NOs:2, 4 or 6. cDNAs generated from alternatively spliced mRNAs, which retain the properties of the Zalpha12 polypeptide are included within the scope of the present invention, as are polypeptides encoded by such cDNAs and mRNAs. Allelic variants and splice variants of these sequences can be cloned by probing cDNA or genomic libraries from different individuals or tissues according to standard procedures known in the art.

Zalpha12 polypeptides that are substantially identical to the polypeptides of SEQ ID NOs:2, 4, 6, 8, 9 or 10 and their orthologs. The term "substantially identical" is used herein to denote polypeptides having 50%, 60%, 70%, 80% and most preferably at least 90%, 95% or 99% sequence identity to the sequences shown in SEQ ID NOs:2, 4, 6, 8, 9 or 10 or their orthologs. Percent sequence identity is

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determined by conventional methods. See, for example, Altschul et al., Bull. Math. Bio. 48: 603-616 (1986) and Henikoff and Henikoff, Proc. Natl. Acad. Sci. USA 89:10915-10919 (1992). Briefly, two amino acid sequences are aligned to optimize the alignment scores using a gap opening penalty of 10, a gap extension penalty of 1, and the "BLOSUM 62" scoring matrix of Henikoff and Henikoff (ibid.) as shown in Table 1 (amino acids are indicated by the standard one-letter codes). The percent identity is then calculated as:

Total number of identical matches

 $- \times 100$

[length of the longer sequence plus the number of gaps introduced into the longer sequence in order to align the two sequences]

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Table 1

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Those skilled in the art appreciate that there are many established algorithms to align two amino acid sequences. The "FASTA" similarity search algorithm of Pearson and Lipman is a suitable protein alignment method . 5 for examining the level of identity shared by an amino acid sequence and the amino acid sequence of a putative variant. The FASTA algorithm is described by Pearson and Lipman, Proc. Nat'l Acad. Sci. USA 85:2444 (1988), and by Pearson, Meth. Enzymol. 183:63 (1990). Briefly, FASTA 10 first characterizes sequence similarity by identifying regions shared by the query sequence (e.g., SEQ ID NO:2) and a test sequence that have either the highest density of identities (if the ktup variable is 1) or pairs of identities (if ktup=2), without considering conservative 15 amino acid substitutions, insertions or deletions. The ten regions with the highest density of identities are then re-scored by comparing the similarity of all paired amino acids using an amino acid substitution matrix, and the ends of the regions are "trimmed" to include only those 20 residues that contribute to the highest score. If there are several regions with scores greater than the "cutoff" value (calculated by a predetermined formula based upon the length of the sequence and the ktup value), then the trimmed initial regions are examined to determine whether 25 the regions can be joined to form an approximate alignment with gaps. Finally, the highest scoring regions of the two amino acid sequences are aligned using a modification of the Needleman-Wunsch-Sellers algorithm (Needleman and Wunsch, J. Mol. Biol. 48:444 (1970); Sellers, SIAM J. 30 Appl. Math. 26:787 (1974), which allows for amino acid insertions and deletions. Illustrative parameters for FASTA analysis are: ktup=1, gap opening penalty=10, gap extension penalty=1, and substitution matrix=BLOSUM62. These parameters can be introduced into a FASTA program by

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modifying the scoring matrix file ("SMATRIX"), as explained in Appendix 2 of Pearson, *Meth. Enzymol.* 183:63 (1990).

FASTA can also be used to determine the sequence identity of nucleic acid molecules using a ratio as disclosed above. For nucleotide sequence comparisons, the ktup value can range between one to six, preferably form four to six.

10 The present invention includes nucleic acid molecules that encode a polypeptide having one or more conservative amino acid changes, compared with the amino acid sequence of SEQ ID NOs:2, 4, 6, 8, 9 or 10. The BLOSUM62 table is an amino acid substitution matrix derived from about 2,000 15 local multiple alignments of protein sequence segments, representing highly conserved regions of more than 500 groups of related proteins [Henikoff and Henikoff, Proc. Nat'l Acad. Sci. USA 89:10915 (1992)]. Accordingly, the BLOSUM62 substitution frequencies can be used to define 20 conservative amino acid substitutions that may be introduced into the amino acid sequences of the present invention. As used herein, the language "conservative amino acid substitution" refers to a substitution represented by a BLOSUM62 value of greater than -1. For 25 example, an amino acid substitution is conservative if the substitution is characterized by a BLOSUM62 value of 0,1,2, or 3. Preferred conservative amino acid substitutions are characterized by a BLOSUM62 value of at least 1 (e.g., 1,2 or 3), while more preferred 30 conservative substitutions are characterized by a BLOSUM62 value of at least 2 (e.g., 2 or 3).

Sequence identity of polynucleotide molecules is determined by similar methods using a ratio as disclosed above.

Variant Zalpha12 polypeptides or substantially homologous Zalpha12 polypeptides are characterized as having one or more amino acid substitutions, deletions or additions. These changes are preferably of a minor nature, that is conservative amino acid substitutions (see Table 2) and other substitutions that do not significantly affect the folding or activity of the polypeptide; small deletions, typically of one to about 30 amino acids; and small amino- or carboxyl-terminal extensions, such as an amino-terminal methionine residue, a small linker peptide of up to about 20-25 residues, or an affinity tag. present invention thus includes polypeptides of from 20 to 30 amino acid residues that comprise a sequence that is at least 90%, preferably at least 95%, and more preferably 99% or more identical to the corresponding region of SEQ 15 ID NOs:2, 4, 6, 8, 9 or 10. Polypeptides comprising affinity tags can further comprise a proteolytic cleavage site between the Zalpha12 polypeptide and the affinity tag. Preferred such sites include thrombin cleavage sites and factor Xa cleavage sites. 20

Table 2

Conservative amino acid substitutions

25

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Basic:

arginine

lysine

histidine

Acidic:

glutamic acid

aspartic acid

Polar:

glutamine

asparagine

24

Table 2 cont.

Hydrophobic: leucine

isoleucine

valine

5 Aromatic: phenylalanine

tryptophan

tyrosine

Small: glycine

alanine

10 serine threonine

methionine

The present invention further provides a variety 15 of other polypeptide fusions [and related multimeric proteins comprising one or more polypeptide fusions]. For example, a Zalpha12 polypeptide can be prepared as a fusion to a dimerizing protein as disclosed in U.S. Patents Nos. 5,155,027 and 5,567,584. Preferred 20 dimerizing proteins in this regard include immunoglobulin constant region domains. Immunoglobulin-Zalpha12 polypeptide fusions can be expressed in genetically engineered cells [to produce a variety of multimeric Zalpha12 analogs]. Auxiliary domains can be fused to 25 Zalpha12 polypeptides to target them to specific cells, tissues, or macromolecules (e.g., collagen). a Zalpha12 polypeptide or protein could be targeted to a predetermined cell type by fusing a Zalpha12 polypeptide to a ligand that specifically binds to a receptor on the 30 surface of the target cell. In this way, polypeptides and proteins can be targeted for therapeutic or diagnostic purposes. A Zalpha12 polypeptide can be fused to two or more moieties, such as an affinity tag for purification and a targeting domain. Polypeptide fusions can also comprise one or more cleavage sites, particularly between

domains. See, Tuan et al., Connective Tissue Research 34:1-9 (1996).

The proteins of the present invention can also comprise non-naturally occurring amino acid residues. 5 Non-naturally occurring amino acids include, without limitation, trans-3-methylproline, 2,4-methanoproline, cis-4-hydroxyproline, trans-4-hydroxyproline, Nmethylglycine, allo-threonine, methylthreonine, hydroxyethylcysteine, hydroxyethylhomocysteine, 10 nitroglutamine, homoglutamine, pipecolic acid, thiazolidine carboxylic acid, dehydroproline, 3- and 4methylproline, 3,3-dimethylproline, tert-leucine, norvaline, 2-azaphenylalanine, 3-azaphenylalanine, 4azaphenylalanine, and 4-fluorophenylalanine. methods are known in the art for incorporating nonnaturally occurring amino acid residues into proteins. For example, an in vitro system can be employed wherein nonsense mutations are suppressed using chemically aminoacylated suppressor tRNAs.

20 Methods for synthesizing amino acids and aminoacylating tRNA are known in the art. Transcription and translation of plasmids containing nonsense mutations is carried out in a cell-free system comprising an E. coli S30 extract and commercially available enzymes and other 25 reagents. Proteins are purified by chromatography. for example, Robertson et al., J. Am. Chem. Soc. 113:2722 (1991); Ellman et al., Methods Enzymol. 202:301 (1991; Chung et al., Science 259:806-809 (1993); and Chung et al., Proc. Natl. Acad. Sci. USA 90:10145-1019 (1993). In 30 a second method, translation is carried out in Xenopus oocytes by microinjection of mutated mRNA and chemically aminoacylated suppressor tRNAs, Turcatti et al., J. Biol. Chem. 271:19991-19998 (1996). Within a third method, E. coli cells are cultured in the absence of a natural amino

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acid that is to be replaced (e.g., phenylalanine) and in the presence of the desired non-naturally occurring amino acid(s) (e.g., 2-azaphenylalanine, 3-azaphenylalanine, 4-azaphenylalanine, or 4-fluorophenylalanine). The non-naturally occurring amino acid is incorporated into the protein in place of its natural counterpart. See, Koide et al., Biochem. 33:7470-7476 (1994). Naturally occurring amino acid residues can be converted to non-naturally occurring species by in vitro chemical modification.

10 Chemical modification can be combined with site-directed mutagenesis to further expand the range of substitutions, Wynn and Richards, Protein Sci. 2:395-403 (1993).

A limited number of non-conservative amino acids, amino acids that are not encoded by the genetic code, non-naturally occurring amino acids, and unnatural amino acids may be substituted for Zalphal2 amino acid residues.

Essential amino acids in the polypeptides of the present invention can be identified according to 20 procedures known in the art, such as site-directed mutagenesis or alanine-scanning mutagenesis, Cunningham and Wells, Science 244: 1081-1085 (1989); Bass et al., Proc. Natl. Acad. Sci. USA 88:4498-502 (1991). latter technique, single alanine mutations are introduced 25 at every residue in the molecule, and the resultant mutant molecules are tested for biological activity as disclosed below to identify amino acid residues that are critical to the activity of the molecule. See also, Hilton et al., J. Biol. Chem. 271:4699-708, 1996. Sites of ligand-receptor 30 interaction can also be determined by physical analysis of structure, as determined by such techniques as nuclear magnetic resonance, crystallography, electron diffraction or photoaffinity labeling, in conjunction with mutation of putative contact site amino acids. See, for example, de 35 Vos et al., Science 255:306-312 (1992); Smith et al., J.

Mol. Biol. 224:899-904 (1992); Wlodaver et al., FEBS Lett. 309:59-64 (1992).

Multiple amino acid substitutions can be made and tested using known methods of mutagenesis and

5 screening, such as those disclosed by Reidhaar-Olson and Sauer, Science 241:53-57 (1988) or Bowie and Sauer, Proc.

Natl. Acad. Sci. USA 86:2152-2156 (1989). Briefly, these authors disclose methods for simultaneously randomizing two or more positions in a polypeptide, selecting for

10 functional polypeptide, and then sequencing the mutagenized polypeptides to determine the spectrum of allowable substitutions at each position. Other methods that can be used include phage display, e.g., Lowman et al., Biochem. 30:10832-10837 (1991); Ladner et al., U.S.

15 Patent No. 5,223,409; Huse, WIPO Publication WO 92/06204) and region-directed mutagenesis, Derbyshire et al., Gene 46:145 (1986); Ner et al., DNA 7:127 (1988).

Variants of the disclosed Zalpha12 DNA and polypeptide sequences can be generated through DNA

20 shuffling as disclosed by Stemmer, Nature 370:389-391,
(1994), Stemmer, Proc. Natl. Acad. Sci. USA 91:10747-10751
(1994) and WIPO Publication WO 97/20078. Briefly, variant DNAs are generated by in vitro homologous recombination by random fragmentation of a parent DNA followed by

reassembly using PCR, resulting in randomly introduced point mutations. This technique can be modified by using a family of parent DNAs, such as allelic variants or DNAs from different species, to introduce additional variability into the process. Selection or screening for the desired activity, followed by additional iterations of mutagenesis and assay provides for rapid "evolution" of sequences by selecting for desirable mutations while simultaneously selecting against detrimental changes.

Mutagenesis methods as disclosed herein can be combined with high-throughput, automated screening methods

to detect activity of cloned, mutagenized polypeptides in host cells. Mutagenized DNA molecules that encode active polypeptides can be recovered from the host cells and rapidly sequenced using modern equipment. These methods allow the rapid determination of the importance of individual amino acid residues in a polypeptide of interest, and can be applied to polypeptides of unknown structure.

Using the methods discussed herein, one of ordinary skill in the art can identify and/or prepare a variety of polypeptide fragments or variants of SEQ ID NOs:2, 4 or 6 or that retain the properties of the wild-type Zalpha12 protein.

For any Zalpha12 polypeptide, including variants
15 and fusion proteins, one of ordinary skill in the art can
readily generate a fully degenerate polynucleotide
sequence encoding that variant using the information set
forth in Tables 1 and 2 above.

20 PROTEIN PRODUCTION

The Zalpha12 polypeptides of the present invention, including full-length polypeptides, biologically active fragments, and fusion polypeptides, can be produced in genetically engineered host cells 25 according to conventional techniques. Suitable host cells are those cell types that can be transformed or transfected with exogenous DNA and grown in culture, and include bacteria, fungal cells, and cultured higher eukaryotic cells. Eukaryotic cells, particularly cultured 30 cells of multicellular organisms, are preferred. Techniques for manipulating cloned DNA molecules and introducing exogenous DNA into a variety of host cells are disclosed by Sambrook et al., Molecular Cloning: A Laboratory Manual, 2nd ed., (Cold Spring Harbor Laboratory 35 Press, Cold Spring Harbor, NY, 1989), and Ausubel et al.,

eds., Current Protocols in Molecular Biology (John Wiley and Sons, Inc., NY, 1987).

In general, a DNA sequence encoding a Zalpha12 polypeptide is operably linked to other genetic elements required for its expression, generally including a transcription promoter and terminator, within an expression vector. The vector will also commonly contain one or more selectable markers and one or more origins of replication, although those skilled in the art will recognize that within certain systems selectable markers

may be provided on separate vectors, and replication of the exogenous DNA may be provided by integration into the host cell genome. Selection of promoters, terminators, selectable markers, vectors and other elements is a matter of routine design within the level of ordinary skill in

the art. Many such elements are described in the literature and are available through commercial suppliers.

To direct a Zalpha12 polypeptide into the secretory pathway of a host cell, a secretory signal sequence (also known as a leader sequence, prepro sequence or pre sequence) is provided in the expression vector. The secretory signal sequence may be that of Zalphal2, or may be derived from another secreted protein (e.g., t-PA)or synthesized de novo. The secretory signal sequence is 25 operably linked to the Zalpha12 DNA sequence, i.e., the two sequences are joined in the correct reading frame and positioned to direct the newly synthesized polypeptide into the secretory pathway of the host cell. Secretory signal sequences are commonly positioned 5' to the DNA sequence encoding the polypeptide of interest, although certain secretory signal sequences may be positioned elsewhere in the DNA sequence of interest (see, e.g., Welch et al., U.S. Patent No. 5,037,743; Holland et al., U.S. Patent No. 5,143,830).

Alternatively, the secretory signal sequence contained in the polypeptides of the present invention is

used to direct other polypeptides into the secretory pathway. The present invention provides for such fusion polypeptides. The secretory signal sequence contained in the fusion polypeptides of the present invention is 5 preferably fused amino-terminally to an additional peptide to direct the additional peptide into the secretory pathway. Such constructs have numerous applications known in the art. For example, these novel secretory signal sequence fusion constructs can direct the secretion of an active component of a normally non-secreted protein, such as a receptor. Such fusions may be used in vivo or in vitro to direct peptides through the secretory pathway.

Cultured mammalian cells are suitable hosts within the present invention. Methods for introducing exogenous DNA into mammalian host cells include calcium phosphate-mediated transfection, Wigler et al., Cell 14:725 (1978); Corsaro and Pearson, Somatic Cell Genetics 7:603 (1981); Graham and Van der Eb, Virology 52:456 (1973), electroporation, Neumann et al., EMBO J. 1:841-845 (1982), DEAE-dextran mediated transfection (Ausubel et al., ibid., and liposome-mediated transfection, Hawley-Nelson et al., Focus 15:73 (1993); Ciccarone et al., Focus 15:80 (1993), and viral vectors, Miller and Rosman, BioTechniques 7:980 (1989); Wang and Finer, Nature Med.

25 2:714 (1996). The production of recombinant polypeptides in cultured mammalian cells is disclosed, for example, by Levinson et al., U.S. Patent No. 4,713,339; Hagen et al., U.S. Patent No. 4,784,950; Palmiter et al., U.S. Patent No. 4,579,821; and Ringold, U.S. Patent No. 4,656,134.
30 Suitable cultured mammalian cells include the COS-1 (ATCC No. CRL 1650), COS-7 (ATCC No. CRL 1651), BHK (ATCC No. CRL 1632), BHK 570 (ATCC No. CRL 10314), 293 (ATCC No. CRL 1573; Graham et al., J. Gen. Virol. 36:59 (1977) and Chinese hamster ovary (e.g. CHO-K1; ATCC No. CCL 61) cell

lines. Additional suitable cell lines are known in the art and available from public depositories such as the American Type Culture Collection, Rockville, Maryland. In general, strong transcription promoters are preferred, such as promoters from SV-40 or cytomegalovirus. See, e.g., U.S. Patent No. 4,956,288. Other suitable promoters include those from metallothionein genes (U.S. Patent Nos. 4,579,821 and 4,601,978) and the adenovirus major late promoter.

10 Drug selection is generally used to select for cultured mammalian cells into which foreign DNA has been inserted. Such cells are commonly referred to as "transfectants". Cells that have been cultured in the presence of the selective agent and are able to pass the 15 gene of interest to their progeny are referred to as "stable transfectants." A preferred selectable marker is a gene encoding resistance to the antibiotic neomycin. Selection is carried out in the presence of a neomycintype drug, such as G-418 or the like. Selection systems can also be used to increase the expression level of the gene of interest, a process referred to as "amplification." Amplification is carried out by culturing transfectants in the presence of a low level of the selective agent and then increasing the amount of selective agent to select for cells that produce high levels of the products of the introduced genes. preferred amplifiable selectable marker is dihydrofolate reductase, which confers resistance to methotrexate. Other drug resistance genes (e.g. hygromycin resistance, 30 multi-drug resistance, puromycin acetyltransferase) can also be used. Alternative markers that introduce an altered phenotype, such as green fluorescent protein, or cell surface proteins such as CD4, CD8, Class I MHC, placental alkaline phosphatase may be used to sort 35 transfected cells from untransfected cells by such means as FACS sorting or magnetic bead separation technology.

Other higher eukaryotic cells can also be used as hosts, including plant cells, insect cells and avian The use of Agrobacterium rhizogenes as a vector for expressing genes in plant cells has been reviewed by 5 Sinkar et al., J. Biosci. (Bangalore) 11:47 (1987). Transformation of insect cells and production of foreign polypeptides therein is disclosed by Guarino et al., U.S. Patent No. 5,162,222 and WIPO publication WO 94/06463. Insect cells can be infected with recombinant baculovirus, 10 commonly derived from Autographa californica nuclear polyhedrosis virus (AcNPV). DNA encoding the Zalpha12 polypeptide is inserted into the baculoviral genome in place of the AcNPV polyhedrin gene coding sequence by one of two methods. The first is the traditional method of 15 homologous DNA recombination between wild-type AcNPV and a transfer vector containing the Zalpha12 flanked by AcNPV Suitable insect cells, e.g. SF9 cells, are sequences. infected with wild-type AcNPV and transfected with a transfer vector comprising a Zalpha12 polynucleotide 20 operably linked to an AcNPV polyhedrin gene promoter, terminator, and flanking sequences. See, King, L.A. and Possee, R.D., The Baculovirus Expression System: A Laboratory Guide, (Chapman & Hall, London); O'Reilly, D.R. et al., Baculovirus Expression Vectors: A Laboratory 25 Manual (Oxford University Press, New York, New York, 1994); and, Richardson, C. D., Ed., Baculovirus Expression Protocols. Methods in Molecular Biology, (Humana Press, Totowa, NJ 1995). Natural recombination within an insect cell will result in a recombinant baculovirus which contains Zalpha12 driven by the polyhedrin promoter. Recombinant viral stocks are made by methods commonly used in the art.

The second method of making recombinant baculovirus utilizes a transposon-based system described by Luckow, V.A, et al., J Virol 67:4566 (1993). This

system is sold in the Bac-to-Bac kit (Life Technologies, Rockville, MD). This system utilizes a transfer vector, $pFastBac1^{TM}$ (Life Technologies) containing a Tn7 transposon to move the DNA encoding the Zalpha12 polypeptide into a 5 baculovirus genome maintained in E. coli as a large plasmid called a "bacmid." The pFastBac1™ transfer vector utilizes the AcNPV polyhedrin promoter to drive the expression of the gene of interest, in this case Zalpha12. However, $pFastBac1^{TM}$ can be modified to a considerable The polyhedrin promoter can be removed and 10 substituted with the baculovirus basic protein promoter (also known as Pcor, p6.9 or MP promoter) which is expressed earlier in the baculovirus infection, and has been shown to be advantageous for expressing secreted 15 proteins. See, Hill-Perkins, M.S. and Possee, R.D., J Gen Virol 71:971 (1990); Bonning, B.C. et al., J Gen Virol 75:1551 (1994); and, Chazenbalk, G.D., and Rapoport, B., JBiol Chem 270:1543 (1995). In such transfer vector constructs, a short or long version of the basic protein 20 promoter can be used. Moreover, transfer vectors can be constructed which replace the native Zalpha12 secretory signal sequences with secretory signal sequences derived from insect proteins. For example, a secretory signal sequence from Ecdysteroid Glucosyltransferase (EGT), honey 25 bee Melittin (Invitrogen, Carlsbad, CA), or baculovirus gp67 (PharMingen, San Diego, CA) can be used in constructs to replace the native Zalpha12 secretory signal sequence. In addition, transfer vectors can include an in-frame fusion with DNA encoding an epitope tag at the C- or N-30 terminus of the expressed Zalpha12 polypeptide, for example, a Glu-Glu epitope tag, Grussenmeyer, T. et al., Proc Natl Acad Sci. 82:7952 (1985). Using a technique known in the art, a transfer vector containing Zalphal2 is transformed into E. coli, and screened for bacmids which contain an interrupted lacZ gene indicative of recombinant

baculovirus. The bacmid DNA containing the recombinant baculovirus genome is isolated, using common techniques, and used to transfect *Spodoptera frugiperda* cells, e.g. Sf9 cells. Recombinant virus that expresses Zalpha12 is subsequently produced. Recombinant viral stocks are made by methods commonly used the art.

5 subsequently produced. Recombinant viral stocks are made by methods commonly used the art. The recombinant virus is used to infect host cells, typically a cell line derived from the fall army worm, Spodoptera frugiperda. See, in general, Glick and 10 Pasternak, Molecular Biotechnology: Principles and Applications of Recombinant DNA (ASM Press, Washington, D.C., 1994). Another suitable cell line is the High FiveO™ cell line (Invitrogen) derived from Trichoplusia ni (U.S. Patent #5,300,435). Commercially available serum-15 free media are used to grow and maintain the cells. Suitable media are Sf900 II $^{ exttt{TM}}$ (Life Technologies) or ESF 921 $^{\text{TM}}$ (Expression Systems) for the Sf9 cells; and ExcellO405™ (JRH Biosciences, Lenexa, KS) or Express FiveO™ (Life Technologies) for the T. ni cells. The cells are 20 grown up from an inoculation density of approximately 2-5 $x\ 10^{5}$ cells to a density of 1-2 $x\ 10^{6}$ cells at which time a recombinant viral stock is added at a multiplicity of infection (MOI) of 0.1 to 10, more typically near 3. recombinant virus-infected cells typically produce the 25 recombinant Zalpha12 polypeptide at 12-72 hours postinfection and secrete it with varying efficiency into the medium. The culture is usually harvested 48 hours postinfection. Centrifugation is used to separate the cells from the medium (supernatant). The supernatant containing 30 the Zalpha12 polypeptide is filtered through micropore filters, usually 0.45 μm pore size. Procedures used are generally described in available laboratory manuals (King, L. A. and Possee, R.D., ibid.; O'Reilly, D.R. et al., ibid.; Richardson, C. D., ibid.). Subsequent purification

of the Zalpha12 polypeptide from the supernatant can be achieved using methods described herein.

Fungal cells, including yeast cells, can also be used within the present invention. Yeast species of 5 particular interest in this regard include Saccharomyces cerevisiae, Pichia pastoris, and Pichia methanolica. Methods for transforming S. cerevisiae cells with exogenous DNA and producing recombinant polypeptides therefrom are disclosed by, for example, Kawasaki, U.S. 10 Patent No. 4,599,311; Kawasaki et al., U.S. Patent No. 4,931,373; Brake, U.S. Patent No. 4,870,008; Welch et al., U.S. Patent No. 5,037,743; and Murray et al., U.S. Patent No. 4,845,075. Transformed cells are selected by phenotype determined by the selectable marker, commonly 15 drug resistance or the ability to grow in the absence of a particular nutrient (e.g., leucine). A preferred vector system for use in Saccharomyces cerevisiae is the POT1 vector system disclosed by Kawasaki et al. (U.S. Patent No. 4,931,373), which allows transformed cells to be 20 selected by growth in glucose-containing media. Suitable promoters and terminators for use in yeast include those from glycolytic enzyme genes (see, e.g., Kawasaki, U.S. Patent No. 4,599,311; Kingsman et al., U.S. Patent No. 4,615,974; and Bitter, U.S. Patent No. 4,977,092) and 25 alcohol dehydrogenase genes. See also U.S. Patents Nos. 4,990,446; 5,063,154; 5,139,936 and 4,661,454. Transformation systems for other yeasts, including Hansenula polymorpha, Schizosaccharomyces pombe, Kluyveromyces lactis, Kluyveromyces fragilis, Ustilago 30 maydis, Pichia pastoris, Pichia methanolica, Pichia guillermondii and Candida maltosa are known in the art. See, for example, Gleeson et al., J. Gen. Microbiol. 132:3459 (1986) and Cregg, U.S. Patent No. 4,882,279.

Aspergillus cells may be utilized according to the methods of McKnight et al., U.S. Patent No. 4,935,349. Methods for transforming Acremonium chrysogenum are disclosed by Sumino et al., U.S. Patent No. 5,162,228. Methods for transforming Neurospora are disclosed by Lambowitz, U.S. Patent No. 4,486,533.

The use of Pichia methanolica as host for the production of recombinant proteins is disclosed in WIPO Publications WO 97/17450, WO 97/17451, WO 98/02536, and WO 10 98/02565. DNA molecules for use in transforming P. methanolica will commonly be prepared as double-stranded, circular plasmids, which are preferably linearized prior to transformation. For polypeptide production in P. methanolica, it is preferred that the promoter and 15 terminator in the plasmid be that of a P. methanolica gene, such as a P. methanolica alcohol utilization gene (AUG1 or AUG2). Other useful promoters include those of the dihydroxyacetone synthase (DHAS), formate dehydrogenase (FMD), and catalase (CAT) genes. 20 facilitate integration of the DNA into the host chromosome, it is preferred to have the entire expression segment of the plasmid flanked at both ends by host DNA sequences. A preferred selectable marker for use in Pichia methanolica is a P. methanolica ADE2 gene, which 25 encodes phosphoribosyl-5-aminoimidazole carboxylase (AIRC; EC 4.1.1.21), which allows ade2 host cells to grow in the absence of adenine. For large-scale, industrial processes where it is desirable to minimize the use of methanol, it is preferred to use host cells in which both methanol 30 utilization genes (AUG1 and AUG2) are deleted. production of secreted proteins, host cells deficient in vacuolar protease genes (PEP4 and PRB1) are preferred. Electroporation is used to facilitate the introduction of a plasmid containing DNA encoding a polypeptide of

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interest into P. methanolica cells. It is preferred to transform P. methanolica cells by electroporation using an exponentially decaying, pulsed electric field having a field strength of from 2.5 to 4.5 kV/cm, preferably about 3.75 kV/cm, and a time constant (t) of from 1 to 40 milliseconds, most preferably about 20 milliseconds.

Prokaryotic host cells, including strains of the bacteria Escherichia coli, Bacillus and other genera are also useful host cells within the present invention. 10 Techniques for transforming these hosts and expressing foreign DNA sequences cloned therein are well known in the art, see, e.g., Sambrook et al., ibid.). When expressing a Zalpha12 polypeptide in bacteria such as E. coli, the polypeptide may be retained in the cytoplasm, typically as 15 insoluble granules, or may be directed to the periplasmic space by a bacterial secretion sequence. In the former case, the cells are lysed, and the granules are recovered and denatured using, for example, guanidine isothiocyanate or urea. The denatured polypeptide can then be refolded 20 and dimerized by diluting the denaturant, such as by dialysis against a solution of urea and a combination of reduced and oxidized glutathione, followed by dialysis against a buffered saline solution. In the latter case, the polypeptide can be recovered from the periplasmic space in a soluble and functional form by disrupting the cells (by, for example, sonication or osmotic shock) to release the contents of the periplasmic space and recovering the protein, thereby obviating the need for denaturation and refolding.

Transformed or transfected host cells are cultured according to conventional procedures in a culture medium containing nutrients and other components required for the growth of the chosen host cells. A variety of suitable media, including defined media and complex media, are known in the art and generally include a carbon source, a nitrogen source, essential amino acids, vitamins

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and minerals. Media may also contain such components as growth factors or serum, as required. The growth medium will generally select for cells containing the exogenously added DNA by, for example, drug selection or deficiency in 5 an essential nutrient which is complemented by the selectable marker carried on the expression vector or cotransfected into the host cell. P. methanolica cells are cultured in a medium comprising adequate sources of carbon, nitrogen and trace nutrients at a temperature of 10 about 25°C to 35°C. Liquid cultures are provided with sufficient aeration by conventional means, such as shaking of small flasks or sparging of fermentors. A preferred culture medium for P. methanolica is YEPD (2% D-glucose, 2% Bacto[™] Peptone (Difco Laboratories, Detroit, MI), 1% 15 Bacto TM yeast extract (Difco Laboratories), 0.004% adenine and 0.006% L-leucine).

Another embodiment of the present invention provides for a peptide or polypeptide comprising an epitope-bearing portion of a Zalpha12 polypeptide of the 20 invention. The epitope of the this polypeptide portion is an immunogenic or antigenic epitope of a polypeptide of the invention. A region of a protein to which an antibody can bind is defined as an "antigenic epitope". See for instance, Geysen, H.M. et al., Proc. Natl. Acad Sci. USA 81:3998-4002 (1984).

As to the selection of peptides or polypeptides bearing an antigenic epitope (i.e., that contain a region of a protein molecule to which an antibody can bind), it is well known in the art that relatively short synthetic 30 peptides that mimic part of a protein sequence are routinely capable of eliciting an antiserum that reacts with the partially mimicked protein. See Sutcliffe, J.G. et al. Science 219:660-666 (1983). Peptides capable of eliciting protein-reactive sera are frequently represented 35 in the primary sequence of a protein, can be characterized by a set of simple chemical rules, and are confined

neither to immunodominant regions of intact proteins (i.e., immunogenic epitopes) nor to the amino or carboxyl terminals. Peptides that are extremely hydrophobic and those of six or fewer residues generally are ineffective at inducing antibodies that bind to the mimicked protein; longer soluble peptides, especially those containing proline residues, usually are effective.

Antigenic epitope-bearing peptides and polypeptides of the invention are therefore useful to 10 raise antibodies, including monoclonal antibodies, that bind specifically to a polypeptide of the invention. Antigenic epitope-bearing peptides and polypeptides of the present invention contain a sequence of at least nine, preferably between 15 to about 30 amino acids contained 15 within the amino acid sequence of a polypeptide of the invention. However, peptides or polypeptides comprising a larger portion of an amino acid sequence of the invention, containing from 30 to 50 amino acids, or any length up to and including the entire amino acid sequence of a 20 polypeptide of the invention, also are useful for inducing antibodies that react with the protein. Preferably, the amino acid sequence of the epitope-bearing peptide is selected to provide substantial solubility in aqueous solvents (i.e., the sequence includes relatively 25 hydrophilic residues and hydrophobic residues are preferably avoided); and sequences containing proline residues are particularly preferred. All of the polypeptides shown in the sequence listing contain antigenic epitopes to be used according to the present 30 invention. The present invention also provides polypeptide fragments or peptides comprising an epitope-bearing portion of a Zalpha12 polypeptide described herein. fragments or peptides may comprise an "immunogenic epitope, " which is a part of a protein that elicits an antibody response when the entire protein is used as an immunogen. Immunogenic epitope-bearing peptides can be identified using standard methods [see, for example,

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Geysen et al., supra. See also U.S. Patent No. 4,708,781 (1987) further describes how to identify a peptide bearing an immunogenic epitope of a desired protein.

Protein Isolation

It is preferred to purify the polypeptides of the present invention to ≥80% purity, more preferably to ≥90% purity, even more preferably ≥95% purity, and

10 particularly preferred is a pharmaceutically pure state, that is greater than 99.9% pure with respect to contaminating macromolecules, particularly other proteins and nucleic acids, and free of infectious and pyrogenic agents. Preferably, a purified polypeptide is

15 substantially free of other polypeptides, particularly other polypeptides of animal origin.

Expressed recombinant Zalpha12 polypeptides (or chimeric Zalpha12 polypeptides) can be purified using fractionation and/or conventional purification methods and media. Ammonium sulfate precipitation and acid or chaotrope extraction may be used for fractionation of samples. Exemplary purification steps may include hydroxyapatite, size exclusion, FPLC and reverse-phase high performance liquid chromatography. Suitable 25 chromatographic media include derivatized dextrans, agarose, cellulose, polyacrylamide, specialty silicas, and the like. PEI, DEAE, QAE and Q derivatives are preferred. Exemplary chromatographic media include those media derivatized with phenyl, butyl, or octyl groups, such as 30 Phenyl-Sepharose FF (Pharmacia), Toyopearl butyl 650 (Toso Haas, Montgomeryville, PA), Octyl-Sepharose (Pharmacia) and the like; or polyacrylic resins, such as Amberchrom CG 71 (Toso Haas) and the like. Suitable solid supports include glass beads, silica-based resins, cellulosic 35 resins, agarose beads, cross-linked agarose beads, polystyrene beads, cross-linked polyacrylamide resins and the like that are insoluble under the conditions in which

they are to be used. These supports may be modified with reactive groups that allow attachment of proteins by amino groups, carboxyl groups, sulfhydryl groups, hydroxyl groups and/or carbohydrate moieties. Examples of coupling chemistries include cyanogen bromide activation, Nhydroxysuccinimide activation, epoxide activation, sulfhydryl activation, hydrazide activation, and carboxyl and amino derivatives for carbodiimide coupling chemistries. These and other solid media are well known and widely used in the art, and are available from commercial suppliers. Methods for binding receptor polypeptides to support media are well known in the art. Selection of a particular method is a matter of routine design and is determined in part by the properties of the 15 chosen support. See, for example, Affinity Chromatography: Principles & Methods (Pharmacia LKB Biotechnology, Uppsala, Sweden, 1988).

The polypeptides of the present invention can be isolated by exploitation of their properties. 20 example, immobilized metal ion adsorption (IMAC) chromatography can be used to purify histidine-rich proteins, including those comprising polyhistidine tags. Briefly, a gel is first charged with divalent metal ions to form a chelate, Sulkowski, Trends in Biochem. 3:1 (1985). Histidine-rich proteins will be adsorbed to this matrix with differing affinities, depending upon the metal ion used, and will be eluted by competitive elution, lowering the pH, or use of strong chelating agents. Other methods of purification include purification of glycosylated proteins by lectin affinity chromatography and ion exchange chromatography. Methods in Enzymol., Vol. 182, "Guide to Protein Purification", M. Deutscher, (ed.),page 529-539 (Acad. Press, San Diego, 1990). Within additional embodiments of the invention, a fusion of the polypeptide of interest and an affinity tag (e.g.,

maltose-binding protein, an immunoglobulin domain) may be constructed to facilitate purification.

Moreover, using methods described in the art, polypeptide fusions, or hybrid Zalpha12 proteins, are constructed using regions or domains of the inventive Zalpha12, Sambrook et al., ibid., Altschul et al., ibid., Picard, Cur. Opin. Biology, 5:511 (1994). These methods allow the determination of the biological importance of larger domains or regions in a polypeptide of interest.

Such hybrids may alter reaction kinetics, binding, constrict or expand the substrate specificity, or alter tissue and cellular localization of a polypeptide, and can be applied to polypeptides of unknown structure.

Fusion proteins can be prepared by methods known 15 to those skilled in the art by preparing each component of the fusion protein and chemically conjugating them. Alternatively, a polynucleotide encoding both components of the fusion protein in the proper reading frame can be generated using known techniques and expressed by the methods described herein. For example, part or all of a domain(s) conferring a biological function may be swapped between Zalpha12 of the present invention with the functionally equivalent domain(s) from another family member. Such domains include, but are not limited to, the 25 secretory signal sequence, conserved, and significant domains or regions in this family. Such fusion proteins would be expected to have a biological functional profile that is the same or similar to polypeptides of the present invention or other known family proteins, depending on the 30 fusion constructed. Moreover, such fusion proteins may exhibit other properties as disclosed herein.

Zalpha12 polypeptides or fragments thereof may also be prepared through chemical synthesis. Zalpha12 polypeptides may be monomers or multimers; glycosylated or non-glycosylated; pegylated or non-pegylated; and may or may not include an initial methionine amino acid residue.

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Chemical Synthesis of Polypeptides
Polypeptides, especially polypeptides of the
present invention can also be synthesized by exclusive

5 solid phase synthesis, partial solid phase methods,
fragment condensation or classical solution synthesis. The
polypeptides are preferably prepared by solid phase
peptide synthesis, for example as described by Merrifield,
J. Am. Chem. Soc. 85:2149 (1963).

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ASSAYS

The activity of molecules of the present invention can be measured using a variety of assays. Of particular interest are changes in steroidogenesis, spermatogenesis, in the testis, LH and FSH production and GnRH in the hypothalamus. Such assays are well known in the art.

Proteins of the present invention are useful for increasing sperm production. Zalpha12 can be measured in 20 vitro using cultured cells or in vivo by administering molecules of the claimed invention to the appropriate animal model. For instance, Zalpha12 transfected (or cotransfected) expression host cells may be embedded in an alginate environment and injected (implanted) into 25 recipient animals. Alginate-poly-L-lysine microencapsulation, permselective membrane encapsulation and diffusion chambers have been described as a means to entrap transfected mammalian cells or primary mammalian These types of non-immunogenic "encapsulations" or 30 microenvironments permit the transfer of nutrients into the microenvironment, and also permit the diffusion of proteins and other macromolecules secreted or released by the captured cells across the environmental barrier to the recipient animal. Most importantly, the capsules or 35 microenvironments mask and shield the foreign, embedded cells from the recipient animal's immune response.

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microenvironments can extend the life of the injected cells from a few hours or days (naked cells) to several weeks (embedded cells).

Alginate threads provide a simple and quick 5 means for generating embedded cells. The materials needed to generate the alginate threads are readily available and relatively inexpensive. Once made, the alginate threads are relatively strong and durable, both in vitro and, based on data obtained using the threads, in vivo. 10 alginate threads are easily manipulable and the methodology is scalable for preparation of numerous threads. In an exemplary procedure, 3% alginate is prepared in sterile H2O, and sterile filtered. to preparation of alginate threads, the alginate solution is again filtered. An approximately 50% cell suspension (containing about 5 x 10^5 to about 5 x 10^7 cells/ml) is mixed with the 3% alginate solution. One ml of the alginate/cell suspension is extruded into a 100 mM sterile filtered CaCl₂ solution over a time period of ~15 min, forming a "thread". The extruded thread is then transferred into a solution of 50 mM $CaCl_2$, and then into a solution of 25 mM CaCl₂. The thread is then rinsed with deionized water before coating the thread by incubating in a 0.01% solution of poly-L-lysine. Finally, the thread is 25 rinsed with Lactated Ringer's Solution and drawn from solution into a syringe barrel (without needle attached). A large bore needle is then attached to the syringe, and the thread is intraperitoneally injected into a recipient

An alternative in vivo approach for assaying proteins of the present invention involves viral delivery systems. Exemplary viruses for this purpose include adenovirus, herpesvirus, vaccinia virus and adenoassociated virus (AAV). Adenovirus, a double-stranded DNA virus, is currently the best studied gene transfer vector for delivery of heterologous nucleic acid (for a review,

in a minimal volume of the Lactated Ringer's Solution.

see T.C. Becker et al., Meth. Cell Biol. 43:161 (1994);
and J.T. Douglas and D.T. Curiel, Science & Medicine 4:44
(1997). The adenovirus system offers several advantages:
adenovirus can (i) accommodate relatively large DNA
inserts; (ii) be grown to high-titer; (iii) infect a broad
range of mammalian cell types; and (iv) be used with a
large number of available vectors containing different
promoters. Also, because adenoviruses are stable in the
bloodstream, they can be administered by intravenous
injection.

By deleting portions of the adenovirus genome, larger inserts (up to 7 kb) of heterologous DNA can be accommodated. These inserts can be incorporated into the viral DNA by direct ligation or by homologous recombination with a co-transfected plasmid. exemplary system, the essential E1 gene has been deleted from the viral vector, and the virus will not replicate unless the El gene is provided by the host cell (the human 293 cell line is exemplary). When intravenously 20 administered to intact animals, adenovirus primarily targets the liver. If the adenoviral delivery system has an El gene deletion, the virus cannot replicate in the host cells. However, the host's tissue (e.g., liver) will express and process (and, if a secretory signal sequence 25 is present, secrete) the heterologous protein. proteins will enter the circulation in the highly vascularized liver, and effects on the infected animal can be determined.

The adenovirus system can also be used for
30 protein production in vitro. By culturing adenovirusinfected non-293 cells under conditions where the cells
are not rapidly dividing, the cells can produce proteins
for extended periods of time. For instance, BHK cells are
grown to confluence in cell factories, then exposed to the
35 adenoviral vector encoding the secreted protein of
interest. The cells are then grown under serum-free

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conditions, which allows infected cells to survive for several weeks without significant cell division.

Alternatively, adenovirus vector infected 293S cells can be grown in suspension culture at relatively high cell density to produce significant amounts of protein (see Garnier et al., Cytotechnol. 15:145 (1994). With either protocol, an expressed, secreted heterologous protein can be repeatedly isolated from the cell culture supernatant. Within the infected 293S cell production protocol, non-secreted proteins may also be effectively obtained.

Agonists and Antagonists

In view of the tissue distribution observed for Zalpha12, agonists (including the natural ligand/
15 substrate/ cofactor/ etc.) and antagonists have enormous potential in both in vitro and in vivo applications.

Compounds identified as Zalpha12 agonists are useful for stimulating the immune system or spermatogenesis. For example, Zalpha12 and agonist compounds are useful as components of defined cell culture media, and may be used alone or in combination with other cytokines and hormones to replace serum that is commonly used in cell culture.

Antagonists

25 Antagonists are also useful as research reagents for characterizing sites of ligand-receptor interaction.

Antagonists of Zalpha12 can also be used to down-regulate inflammation as discussed in more further detail below.

Inhibitors of Zalpha12 activity (Zalpha12 antagonists)

30 include anti-Zalpha12 antibodies and soluble Zalpha12 receptors, as well as other peptidic and non-peptidic agents (including ribozymes).

Zalpha12 can also be used to identify inhibitors (antagonists) of its activity. Test compounds are added to the assays disclosed herein to identify compounds that inhibit the activity of Zalpha12. In addition to those assays disclosed herein, samples can be tested for

inhibition of Zalphal2 activity within a variety of assays designed to measure receptor binding or the stimulation/inhibition of Zalpha12-dependent cellular responses. For example, Zalpha12-responsive cell lines 5 can be transfected with a reporter gene construct that is responsive to a Zalpha12-stimulated cellular pathway. Reporter gene constructs of this type are known in the art, and will generally comprise a Zalpha12-DNA response element operably linked to a gene encoding a protein which 10 can be assayed, such as luciferase. DNA response elements can include, but are not limited to, cyclic AMP response elements (CRE), hormone response elements (HRE) insulin response element (IRE), Nasrin et al., Proc. Natl. Acad. Sci. USA 87:5273 (1990) and serum response elements (SRE) 15 (Shaw et al. Cell 56: 563 (1989). Cyclic AMP response elements are reviewed in Roestler et al., J. Biol. Chem. 263 (19):9063 (1988) and Habener, Molec. Endocrinol. 4 (8):1087 (1990). Hormone response elements are reviewed in Beato, Cell 56:335 (1989). Candidate compounds, 20 solutions, mixtures or extracts are tested for the ability to inhibit the activity of Zalphal2 on the target cells as evidenced by a decrease in Zalpha12 stimulation of reporter gene expression. Assays of this type will detect compounds that directly block Zalpha12 binding to cell-25 surface receptors, as well as compounds that block processes in the cellular pathway subsequent to receptorligand binding. In the alternative, compounds or other samples can be tested for direct blocking of Zalpha12 binding to receptor using Zalpha12 tagged with a 30 detectable label (e.g., 125I, biotin, horseradish peroxidase, FITC, and the like). Within assays of this type, the ability of a test sample to inhibit the binding of labeled Zalpha12 to the receptor is indicative of inhibitory activity, which can be confirmed through

35 secondary assays. Receptors used within binding assays

may be cellular receptors or isolated, immobilized receptors.

A Zalpha12 polypeptide can be expressed as a fusion with an immunoglobulin heavy chain constant region, typically an F_C fragment, which contains two constant region domains and lacks the variable region. Methods for preparing such fusions are disclosed in U.S. Patents Nos. 5,155,027 and 5,567,584. Such fusions are typically secreted as multimeric molecules wherein the Fc portions are disulfide bonded to each other and two non-Ig polypeptides are arrayed in closed proximity to each other. Fusions of this type can be used to affinity purify the ligand. For use in assays, the chimeras are bound to a support via the F_C region and used in an ELISA format.

A Zalpha12 ligand-binding polypeptide can also be used for purification of ligand. The polypeptide is immobilized on a solid support, such as beads of agarose, cross-linked agarose, glass, cellulosic resins, silica-20 based resins, polystyrene, cross-linked polyacrylamide, or like materials that are stable under the conditions of Methods for linking polypeptides to solid supports are known in the art, and include amine chemistry, cyanogen bromide activation, N-hydroxysuccinimide 25 activation, epoxide activation, sulfhydryl activation, and hydrazide activation. The resulting medium will generally be configured in the form of a column, and fluids containing ligand are passed through the column one or more times to allow ligand to bind to the receptor 30 polypeptide. The ligand is then eluted using changes in salt concentration, chaotropic agents (guanidine HCl), or pH to disrupt ligand-receptor binding.

An assay system that uses a ligand-binding receptor (or an antibody, one member of a complement/

anti-complement pair) or a binding fragment thereof, and a commercially available biosensor instrument (BIAcore, Pharmacia Biosensor, Piscataway, NJ) may be advantageously

employed. Such receptor, antibody, member of a complement/anti-complement pair or fragment is immobilized onto the surface of a receptor chip. Use of this instrument is disclosed by Karlsson, J. Immunol. Methods 5 145:229 (1991) and Cunningham and Wells, J. Mol. Biol. 234:554 (1993). A receptor, antibody, member or fragment is covalently attached, using amine or sulfhydryl chemistry, to dextran fibers that are attached to gold film within the flow cell. A test sample is passed 10 through the cell. If a ligand, epitope, or opposite member of the complement/anti-complement pair is present in the sample, it will bind to the immobilized receptor, antibody or member, respectively, causing a change in the refractive index of the medium, which is detected as a change in surface plasmon resonance of the gold film. This system allows the determination of on- and off-rates, from which binding affinity can be calculated, and assessment of stoichiometry of binding.

Ligand-binding receptor polypeptides can also be used within other assay systems known in the art. Such systems include Scatchard analysis for determination of binding affinity, Scatchard, Ann. NY Acad. Sci. 51: 660 (1949) and calorimetric assays, Cunningham et al., Science 253:545 (1991); Cunningham et al., Science 245:821 (1991).

Zalpha12 polypeptides can also be used to prepare antibodies that specifically bind to Zalpha12 epitopes, peptides or polypeptides. The Zalpha12 polypeptide or a fragment thereof serves as an antigen (immunogen) to inoculate an animal and elicit an immune response. Suitable antigens would be the Zalpha12 polypeptides encoded by SEQ ID NOs: 2, 4, 6, 8-10, 18-20, 25-30 and 35-44. Antibodies generated from this immune response can be isolated and purified as described herein. Methods for preparing and isolating polyclonal and monoclonal antibodies are well known in the art. See, for example, Current Protocols in Immunology, Cooligan, et al.

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(eds.), National Institutes of Health, (John Wiley and Sons, Inc., 1995); Sambrook et al., Molecular Cloning: A Laboratory Manual, Second Edition (Cold Spring Harbor, NY, 1989); and Hurrell, J. G. R., Ed., Monoclonal Hybridoma 5 Antibodies: Techniques and Applications (CRC Press, Inc., Boca Raton, FL, 1982).

As would be evident to one of ordinary skill in the art, polyclonal antibodies can be generated from inoculating a variety of warm-blooded animals such as 10 horses, cows, goats, sheep, dogs, chickens, rabbits, mice, and rats with a Zalpha12 polypeptide or a fragment thereof. The immunogenicity of a Zalpha12 polypeptide may be increased through the use of an adjuvant, such as alum (aluminum hydroxide) or Freund's complete or incomplete 15 adjuvant. Polypeptides useful for immunization also include fusion polypeptides, such as fusions of Zalpha12 or a portion thereof with an immunoglobulin polypeptide or with maltose binding protein. The polypeptide immunogen may be a full-length molecule or a portion thereof. 20 the polypeptide portion is "hapten-like", such portion may be advantageously joined or linked to a macromolecular carrier (such as keyhole limpet hemocyanin (KLH), bovine serum albumin (BSA) or tetanus toxoid) for immunization.

As used herein, the term "antibodies" includes polyclonal antibodies, affinity-purified polyclonal antibodies, monoclonal antibodies, and antigen-binding fragments, such as F(ab')₂, Fab proteolytic fragments, Genetically engineered intact antibodies or fragments, such as chimeric antibodies, Fv fragments, single chain antibodies and the like, as well as synthetic antigen-binding peptides and polypeptides. Non-human antibodies may be humanized by grafting non-human CDRs onto human framework and constant regions, or by incorporating the entire non-human variable domains (optionally "cloaking" them with a human-like surface by replacement of exposed residues, wherein the result is a "veneered" antibody).

In some instances, humanized antibodies may retain nonhuman residues within the human variable region framework domains to enhance proper binding characteristics. Through humanizing antibodies, biological half-life may be increased, and the potential for adverse immune reactions upon administration to humans is reduced.

Alternative techniques for generating or selecting antibodies useful herein include in vitro exposure of lymphocytes to Zalpha12 protein or peptide, and selection of antibody display libraries in phage or similar vectors (for instance, through use of immobilized or labeled Zalpha12 protein or peptide). Genes encoding polypeptides having potential Zalpha12 polypeptide binding domains can be obtained by screening random peptide libraries displayed on phage (phage display) or on bacteria, such as E. coli. Nucleotide sequences encoding the polypeptides can be obtained in a number of ways, such as through random mutagenesis and random polynucleotide synthesis. These random peptide display libraries can be used to screen for peptides which interact with a known target which can be a protein or polypeptide, such as a ligand or receptor, a biological or synthetic macromolecule, or organic or inorganic substances. Techniques for creating and screening such random peptide 25 display libraries are known in the art (Ladner et al., US Patent NO. 5,223,409; Ladner et al., US Patent NO. 4,946,778; Ladner et al., US Patent NO. 5,403,484 and Ladner et al., US Patent NO. 5,571,698) and random peptide display libraries and kits for screening such libraries 30 are available commercially, for instance from Clontech (Palo Alto, CA), Invitrogen Inc. (San Diego, CA), New England Biolabs, Inc. (Beverly, MA) and Pharmacia LKB Biotechnology Inc. (Piscataway, NJ). Random peptide display libraries can be screened using the Zalpha12 sequences disclosed herein to identify proteins which bind to Zalpha12. These "binding proteins" which interact

with Zalpha12 polypeptides can be used for tagging cells; for isolating homolog polypeptides by affinity purification; they can be directly or indirectly conjugated to drugs, toxins, radionuclides and the like.

5 These binding proteins can also be used in analytical methods such as for screening expression libraries and neutralizing activity. The binding proteins can also be used for diagnostic assays for determining circulating levels of polypeptides; for detecting or quantitating soluble polypeptides as marker of underlying pathology or disease. These binding proteins can also act as Zalpha12 "antagonists" to block Zalpha12 binding and signal transduction in vitro and in vivo.

Antibodies are determined to be specifically

binding if: (1) they exhibit a threshold level of binding
activity, and (2) they do not significantly cross-react
with related polypeptide molecules. First, antibodies
herein specifically bind if they bind to a Zalpha12
polypeptide, peptide or epitope with a binding affinity

(Ka) of 10⁶ M⁻¹ or greater, preferably 10⁷ M⁻¹ or greater,
more preferably 10⁸ M⁻¹ or greater, and most preferably
10⁹ M⁻¹ or greater. The binding affinity of an antibody
can be readily determined by one of ordinary skill in the
art, for example, by Scatchard analysis.

Second, antibodies are determined to specifically bind if they do not significantly cross-react with related polypeptides. Antibodies do not significantly cross-react with related polypeptide molecules, for example, if they detect Zalpha12 but not known related polypeptides using a standard Western blot analysis (Ausubel et al., ibid.). Examples of known related polypeptides are orthologs, proteins from the same species that are members of a protein family (e.g. IL-16), Zalpha12 polypeptides, and non-human Zalpha12. Moreover, antibodies may be "screened against" known related

polypeptides to isolate a population that specifically binds to the inventive polypeptides. For example, antibodies raised to Zalpha12 are adsorbed to related polypeptides adhered to insoluble matrix; antibodies 5 specific to Zalpha12 will flow through the matrix under the proper buffer conditions. Such screening allows isolation of polyclonal and monoclonal antibodies noncrossreactive to closely related polypeptides, Antibodies: A Laboratory Manual, Harlow and Lane (eds.) (Cold Spring 10 Harbor Laboratory Press, 1988); Current Protocols in Immunology, Cooligan, et al. (eds.), National Institutes of Health (John Wiley and Sons, Inc., 1995). Screening and isolation of specific antibodies is well known in the art. See, Fundamental Immunology, Paul (eds.) (Raven 15 Press, 1993); Getzoff et al., Adv. in Immunol. 43: 1-98 (1988); Monoclonal Antibodies: Principles and Practice, Goding, J.W. (eds.), (Academic Press Ltd., 1996); Benjamin et al., Ann. Rev. Immunol. 2: 67-101 (1984).

A variety of assays known to those skilled in
the art can be utilized to detect antibodies which
specifically bind to Zalpha12 proteins or peptides.
Exemplary assays are described in detail in Antibodies: A
Laboratory Manual, Harlow and Lane (Eds.) (Cold Spring
Harbor Laboratory Press, 1988). Representative examples
of such assays include: concurrent immunoelectrophoresis,
radioimmunoassay, radioimmuno-precipitation, enzyme-linked
immunosorbent assay (ELISA), dot blot or Western blot
assay, inhibition or competition assay, and sandwich
assay. In addition, antibodies can be screened for
binding to wild-type versus mutant Zalpha12 protein or
polypeptide.

Antibodies to Zalpha12 may be used for tagging cells that express Zalpha12; for isolating Zalpha12 by affinity purification; for diagnostic assays for determining circulating levels of Zalpha12 polypeptides;

for detecting or quantitating soluble Zalpha12 as marker of underlying pathology or disease; in analytical methods employing FACS; for screening expression libraries; for generating anti-idiotypic antibodies; and as neutralizing 5 antibodies or as antagonists to block Zalpha12 in vitro and in vivo. Suitable direct tags or labels include radionuclides, enzymes, substrates, cofactors, inhibitors, fluorescent markers, chemiluminescent markers, magnetic particles and the like; indirect tags or labels may 10 feature use of biotin-avidin or other complement/anticomplement pairs as intermediates. Antibodies herein may also be directly or indirectly conjugated to drugs, toxins, radionuclides and the like, and these conjugates used for in vivo diagnostic or therapeutic applications. 15 Moreover, antibodies to Zalpha12 or fragments thereof may be used in vitro to detect denatured Zalpha12 or fragments thereof in assays, for example, Western Blots or other

20 BIOACTIVE CONJUGATES:

assays known in the art.

Antibodies or polypeptides herein can also be directly or indirectly conjugated to drugs, toxins, radionuclides and the like, and these conjugates used for in vivo diagnostic or therapeutic applications. For instance, polypeptides or antibodies of the present invention can be used to identify or treat tissues or organs that express a corresponding anti-complementary molecule (receptor or antigen, respectively, for instance). More specifically, Zalpha12 polypeptides or anti-Zalpha12 antibodies, or bioactive fragments or portions thereof, can be coupled to detectable or cytotoxic molecules and delivered to a mammal having cells, tissues or organs that express the anti-complementary molecule.

35 Suitable detectable molecules may be directly or indirectly attached to the polypeptide or antibody, and

include radionuclides, enzymes, substrates, cofactors, inhibitors, fluorescent markers, chemiluminescent markers, magnetic particles and the like. Suitable cytotoxic molecules may be directly or indirectly attached to the 5 polypeptide or antibody, and include bacterial or plant toxins (for instance, diphtheria toxin, Pseudomonas exotoxin, ricin, abrin and the like), as well as therapeutic radionuclides, such as iodine-131, rhenium-188 or yttrium-90 (either directly attached to the polypeptide 10 or antibody, or indirectly attached through means of a chelating moiety, for instance). Polypeptides or antibodies may also be conjugated to cytotoxic drugs, such as adriamycin. For indirect attachment of a detectable or cytotoxic molecule, the detectable or cytotoxic molecule can be conjugated with a member of a complementary/ anticomplementary pair, where the other member is bound to the polypeptide or antibody portion. For these purposes, biotin/streptavidin is an exemplary complementary/ anticomplementary pair.

20 In another embodiment, polypeptide-toxin fusion proteins or antibody-toxin fusion proteins can be used for targeted cell or tissue inhibition or ablation (for instance, to treat cancer cells or tissues). Alternatively, if the polypeptide has multiple functional 25 domains (i.e., an activation domain or a ligand binding domain, plus a targeting domain), a fusion protein including only the targeting domain may be suitable for directing a detectable molecule, a cytotoxic molecule or a complementary molecule to a cell or tissue type of 30 interest. In instances where the domain only fusion protein includes a complementary molecule, the anticomplementary molecule can be conjugated to a detectable or cytotoxic molecule. Such domain-complementary molecule fusion proteins thus represent a generic targeting vehicle 35 for cell/tissue-specific delivery of generic anticomplementary-detectable/ cytotoxic molecule conjugates.

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In another embodiment, Zalpha12-cytokine fusion proteins or antibody-cytokine fusion proteins can be used for enhancing in vivo killing of target tissues (for example, blood and bone marrow cancers), if the Zalpha12 5 polypeptide or anti-Zalpha12 antibody targets the hyperproliferative blood or bone marrow cell. See, generally, Hornick et al., Blood 89:4437 (1997). The described fusion proteins enable targeting of a cytokine to a desired site of action, thereby providing an elevated 10 local concentration of cytokine. Suitable Zalpha12 polypeptides or anti-Zalpha12 antibodies target an undesirable cell or tissue (i.e., a tumor or a leukemia), and the fused cytokine mediated improved target cell lysis by effector cells. Suitable cytokines for this purpose 15 include interleukin 2 and granulocyte-macrophage colonystimulating factor (GM-CSF); for instance.

In yet another embodiment, if the Zalpha12 polypeptide or anti-Zalpha12 antibody targets vascular cells or tissues, such polypeptide or antibody may be 20 conjugated with a radionuclide, and particularly with a beta-emitting radionuclide, to reduce restenosis. Such therapeutic approach poses less danger to clinicians who administer the radioactive therapy. For instance, iridium-192 impregnated ribbons placed into stented 25 vessels of patients until the required radiation dose was delivered showed decreased tissue growth in the vessel and greater luminal diameter than the control group, which received placebo ribbons. Further, revascularisation and stent thrombosis were significantly lower in the treatment 30 group. Similar results are predicted with targeting of a bioactive conjugate containing a radionuclide, as described herein.

The bioactive polypeptide or antibody conjugates described herein can be delivered intravenously,

intraarterially or intraductally, or may be introduced locally at the intended site of action.

USES OF POLYNUCLEOTIDE/POLYPEPTIDE:

Molecules of the present invention can be used to identify and isolate receptors involved in 5 spermatogenesis, steroidogenesis, testicular differentiation and regulatory control of the hypothalamic-pituitary-gonadal axis or receptors of the immune system. For example, proteins and peptides of the present invention can be immobilized on a column and 10 membrane preparations run over the column, Immobilized Affinity Ligand Techniques, Hermanson et al., eds., pp.195-202 (Academic Press, San Diego, CA, 1992,). Proteins and peptides can also be radiolabeled, Methods in Enzymol., vol. 182, "Guide to Protein Purification", M. 15 Deutscher, ed., pp 721-737 (Acad. Press, San Diego, 1990) or photoaffinity labeled, Brunner et al., Ann. Rev. Biochem. 62:483-514 (1993) and Fedan et al., Biochem. Pharmacol. 33:1167 (1984) and specific cell-surface proteins can be identified.

The molecules of the present invention will be useful for testing disorders of the reproductive system and immunological systems.

GENE THERAPY:

Polynucleotides encoding Zalpha12 polypeptides are useful within gene therapy applications where it is desired to increase or inhibit Zalpha12 activity. If a mammal has a mutated or absent Zalpha12 gene, the Zalpha12 gene can be introduced into the cells of the mammal. In one embodiment, a gene encoding a Zalpha12 polypeptide is introduced in vivo in a viral vector. Such vectors include an attenuated or defective DNA virus, such as, but not limited to, herpes simplex virus (HSV), papillomavirus, Epstein Barr virus (EBV), adenovirus, adeno-associated virus (AAV), and the like. Defective

viruses, which entirely or almost entirely lack viral genes, are preferred. A defective virus is not infective after introduction into a cell. Use of defective viral vectors allows for administration to cells in a specific, localized area, without concern that the vector can infect other cells. Examples of particular vectors include, but are not limited to, a defective herpes simplex virus 1 (HSV1) vector, Kaplitt et al., Molec. Cell. Neurosci. 2:320 (1991); an attenuated adenovirus vector, such as the vector described by Stratford-Perricaudet et al., J. Clin. Invest. 90:626 (1992); and a defective adeno-associated virus vector, Samulski et al., J. Virol. 61:3096 (1987); Samulski et al., J. Virol. 63:3822 (1989).

In another embodiment, a Zalpha12 gene can be introduced in a retroviral vector, e.g., as described in Anderson et al., U.S. Patent No. 5,399,346; Mann et al. Cell 33:153, 1983; Temin et al., U.S. Patent No. 4,650,764; Temin et al., U.S. Patent No. 4,980,289; Markowitz et al., J. Virol. 62:1120 (1988); Temin et al.,

- 20 U.S. Patent No. 5,124,263; International Patent Publication No. WO 95/07358, published March 16, 1995 by Dougherty et al.; and Kuo et al., Blood 82:845 (1993). Alternatively, the vector can be introduced by lipofection in vivo using liposomes. Synthetic cationic lipids can be
- used to prepare liposomes for in vivo transfection of a gene encoding a marker, Felgner et al., Proc. Natl. Acad. Sci. USA 84:7413 (1987); Mackey et al., Proc. Natl. Acad. Sci. USA 85:8027 (1988). The use of lipofection to introduce exogenous genes into specific organs in vivo has
- 30 certain practical advantages. Molecular targeting of liposomes to specific cells represents one area of benefit. More particularly, directing transfection to particular cells represents one area of benefit. For

instance, directing transfection to particular cell types would be particularly advantageous in a tissue with cellular heterogeneity, such as the pancreas, liver, kidney, and brain. Lipids may be chemically coupled to other molecules for the purpose of targeting. Targeted peptides (e.g., hormones or neurotransmitters), proteins such as antibodies, or non-peptide molecules can be coupled to liposomes chemically.

It is possible to remove the target cells from
the body; to introduce the vector as a naked DNA plasmid;
and then to re-implant the transformed cells into the
body. Naked DNA vectors for gene therapy can be
introduced into the desired host cells by methods known in
the art, e.g., transfection, electroporation,

- microinjection, transduction, cell fusion, DEAE dextran, calcium phosphate precipitation, use of a gene gun or use of a DNA vector transporter. See, e.g., Wu et al., J. Biol. Chem. 267:963 (1992); Wu et al., J. Biol. Chem. 263:14621-4, 1988.
- Antisense methodology can be used to inhibit Zalpha12 gene transcription, such as to inhibit cell proliferation in vivo. Polynucleotides that are complementary to a segment of a Zalpha12-encoding polynucleotide (e.g., a polynucleotide as set froth in SEQ
- 25 ID NO:1) are designed to bind to Zalpha12-encoding mRNA and to inhibit translation of such mRNA. Such antisense polynucleotides are used to inhibit expression of Zalpha12 polypeptide-encoding genes in cell culture or in a subject.
- The present invention also provides reagents which will find use in diagnostic applications. For example, the Zalpha12 gene, a probe comprising Zalpha12 DNA or RNA or a subsequence thereof can be used to determine if the Zalpha12 gene is present on chromosome
- 35 22q13.1 or if a mutation has occurred. Detectable chromosomal aberrations at the Zalpha12 gene locus

include, but are not limited to, aneuploidy, gene copy number changes, insertions, deletions, restriction site changes and rearrangements. Such aberrations can be detected using polynucleotides of the present invention by 5 employing molecular genetic techniques, such as restriction fragment length polymorphism (RFLP) analysis, short tandem repeat (STR) analysis employing PCR techniques, and other genetic linkage analysis techniques known in the art (Sambrook et al., ibid.; Ausubel et. al., ibid.; Marian, Chest 108:255 (1995).

Transgenic mice, engineered to express the Zalpha12 gene, and mice that exhibit a complete absence of Zalpha12 gene function, referred to as "knockout mice", Snouwaert et al., Science 257:1083 (1992), may also be 15 generated, Lowell et al., Nature 366:740-42 (1993). mice may be employed to study the Zalpha12 gene and the protein encoded thereby in an in vivo system.

CHROMOSOMAL LOCALIZATION:

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20 Zalpha12 has been mapped to chromosome 22q13.1. Radiation hybrid mapping is a somatic cell genetic technique developed for constructing high-resolution, contiguous maps of mammalian chromosomes (Cox et al., Science 250:245 (1990). Partial or full knowledge of a 25 gene's sequence allows one to design PCR primers suitable for use with chromosomal radiation hybrid mapping panels. Radiation hybrid mapping panels are commercially available which cover the entire human genome, such as the Stanford G3 RH Panel and the GeneBridge 4 RH Panel (Research 30 Genetics, Inc., Huntsville, AL). These panels enable rapid, PCR-based chromosomal localizations and ordering of genes, sequence-tagged sites (STSs), and other nonpolymorphic and polymorphic markers within a region of interest. This includes establishing directly 35 proportional physical distances between newly discovered

genes of interest and previously mapped markers. The precise knowledge of a gene's position can be useful for a number of purposes, including: 1) determining if a sequence is part of an existing contig and obtaining additional surrounding genetic sequences in various forms, such as YACs, BACs or cDNA clones; 2) providing a possible candidate gene for an inheritable disease which shows linkage to the same chromosomal region; and 3) cross-referencing model organisms, such as mouse, which may aid in determining what function a particular gene might have.

Sequence tagged sites (STSs) can also be used independently for chromosomal localization. An STS is a DNA sequence that is unique in the human genome and can be used as a reference point for a particular chromosome or region of a chromosome. An STS is defined by a pair of oligonucleotide primers that are used in a polymerase chain reaction to specifically detect this site in the presence of all other genomic sequences. Since STSs are based solely on DNA sequence they can be completely described within an electronic database, for example, Database of Sequence Tagged Sites (dbSTS), GenBank, (National Center for Biological Information, National Institutes of Health, Bethesda, MD

within these short genomic landmark STS sequences.

For pharmaceutical use, the proteins of the present invention are formulated for parenteral,

particularly intravenous or subcutaneous, delivery according to conventional methods. Intravenous administration will be by bolus injection or infusion over a typical period of one to several hours. In general, pharmaceutical formulations will include a Zalpha12

protein in combination with a pharmaceutically acceptable vehicle, such as saline, buffered saline, 5% dextrose in water or the like. Formulations may further include one

gene sequence of interest for the mapping data contained

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or more excipients, preservatives, solubilizers, buffering agents, albumin to prevent protein loss on vial surfaces, Methods of formulation are well known in the art and are disclosed, for example, in Remington: The Science and 5 Practice of Pharmacy, Gennaro, ed., (Mack Publishing Co., Easton, PA, 19th ed., 1995). Therapeutic doses will generally be in the range of 0.1 to 100 $\mu g/kg$ of patient weight per day, preferably 0.5-20 mg/kg per day, with the exact dose determined by the clinician according to 10 accepted standards, taking into account the nature and severity of the condition to be treated, patient traits, Determination of dose is within the level of ordinary skill in the art. The proteins may be administered for acute treatment, over one week or less, 15 often over a period of one to three days or may be used in chronic treatment, over several months or years. Administration of the protein can be subcutaneous, intraperitoneal or rectal depending on the disease to be treated.

20

Tissue Expression and Use

Zalpha12 represents a novel polypeptide with a putative signal peptide leader sequence and alpha helical structure. Several putative isoforms have been

25 identified. Therefore this gene may encode a secreted polypeptide with secondary structure indicating it is a member of the four helix bundle cytokine family. Alternatively, this polypeptide may have other activities associated with other biological functions including:

30 enzymatic activity, association with the cell membrane, or function as a carrier protein.

Northern blot analysis detects transcripts for zalpha12 in spleen, thymus, testis, small intestine, colon, PBL, stomach, lymph node, trachea, and bone marrow.

Many of these organs have important immunological function or contain cells which play a role in the immune system.

Zalpha12 is also expressed in CD4+ enriched peripheral T cells and to a lesser degree in CD8+ T cells. very weak expression in CD19+ B cells suggesting that Zalpha12 has expression in T lymphoid lineage cells and 5 little if any in B lymphoid lineage cells. elevated expression of zalpha12 in RNA derived from a 7 day human mixed lymphocyte reaction. This suggests that zalpha12 expression is regulated and increases after T cell activation.

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Use of Zalpha12

Zalpha12 can be administered to an immunocompromised mammal, preferably a human, such as cancer patients who have undergone chemotherapy, AIDS patients and the 15 elderly. This will stimulate their immune systems. Zalpha12 can also be used as a vaccine adjuvant to be administered before, with or after the administration of a vaccine. Zalpha12 may also be administered to stimulate the immune system to attack tumors.

20

Use of Antagonists of Zalpha12

An antagonist to Zalpha12, such as an antibody, soluble receptor or small molecule antagonist can be administered to a mammal, preferably a human, to alleviate an inflammatory response. Antagonists, such as antibodies, 25 to Zalpha12 can be used to treat patients having inflammatory related diseases such arteriosclerotic heart disease [see Paulsson, G. et al., Arterioscler Thromb. Vasc. Biol., 20:10-17 (2000)], inflammatory bowel disease, Crohn's disease, rheumatoid arthritis and pancreatitis.

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The invention is further illustrated by the following non-limiting examples.

Example 1

Cloning of Zalpha12

35 Zalpha12 was discovered by using the Expressed Sequence Tag of SEQ ID NO:7 as a probe in a spleen cDNA library. The library was constructed using 1 microgram

(μg) of polyA RNA isolated from the spleen tissue of a 2-year-old Hispanic male, who died from cerebral anoxia. cDNA synthesis was initiated using a NotI-oligo(dT) primer. Double-stranded cDNA was blunted, ligated to EcoRI adaptors, digested with NotI, size-selected, and cloned into the NotI and EcoRI sites of a vector. The sequenced clone resulted in the full-length DNA and polypeptide sequences of SEQ ID NOs: 1 and 2.

The EST of SEQ ID NO: 17 resulted in the 10 discovery of a second clone. The clone was sequence and found to have an unspliced intron. Oligonucleotide primers SEQ ID NOs: 11 and 12 were designed to amplify the fulllength sequence without the intron sequence. The PCR was performed using lymph node MARATHON READY® cDNA (Clontech, 15 Palo Alto, CA) as template. An 823 bp was cut out and gel purified using the QIAQUICK® gel extraction kit (Qiagen, Santa Clarita, CA). The excised PCR fragment was sequenced and did not contain the intron sequence. The next step was to see if the clone of SEQ ID NO:17 had another initiation 20 codon upstream. A RACE® reaction using lymph node cDNA performed to obtain 5' sequence. Oligonucleotide primers SEQ ID NOs:13 and 14 were designed for 5' RACE® reactions and nested RACE®. The RACE® reactions revealed an additional initiation codon upstream. The actual fulllength was obtained through PCR using primers SEQ ID NOs:15 and 16 which flanked the coding sequence. Six bands were amplified and each was gel purified using the QIAQUICK® gel extraction kit and subcloned into plasmid pCR2.1TOPO vector using the TOPO TA® cloning (Invitrogen, Carlsbad, CA). The clones were sequenced and 30 resulted in the full-length sequences of SEQ ID NOs: 3, 4, 5 and 6.

Thus, three variants were discovered, namely SEQ ID NO:1 and 2, SEQ ID NO:3 and 4, and SEQ ID NOs: 5 and 6.

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Example 2

Northern Blot Analysis of Zalpha12

A northern blot analysis was carried out by standard techniques using the polynucleotides of SEQ ID NOs: 1 and 3. The results indicate that Zalpha12 is expressed in lymph nodes, activated mixed lymphocytes, spleen, thymus, testis, small intestines, human aortic endothelial cells, smooth muscle, kidney, mast cells, eosinophils, tonsils, pancreas, colon, peripheral blood lymphocytes (PBL), stomach, trachea, T-cells including CD4 and CD8 cells, and bone marrow.

Zalpha12 is also expressed by the following cell lines: HL60 (ATCC 45500), an acute promyelocytic leukemia cell line; Jurkat cells (ATCC TIB-152) a T lymphocyte from acute T cell leukemia; MOLT-4 cells (ATCC CRL-1582) a T lymphoblast from acute lymphoblastic leukemia; and HuT 78 cells (ATCC TIB-161) a cutaneous T lymphocyte from a lymphoma.

20 From these data, it can be concluded that Zalpha12 is a cytokine involved in the inflammation cascade.

Antagonists of Zalpha12 can be used to alleviate inflammation related to a number of diseases such as arteriosclerotic heart disease, cardiovascular disease, rheumatoid arthritis, inflammatory bowel disease, and Crohn's disease.

Example 3

Zalpha12 Anti-peptide Antibodies

30

Polyclonal anti-peptide antibodies were prepared by immunizing two female New Zealand white rabbits with the peptide huzalpha12X1-1

AQQHKGSLQKDPLLSQACVGCLEALLDYLDAR (SEQ ID NO:41) or the peptide from SEQ ID NO:4 PLPATKDTVLAPLRMSQVRSLVIGLQNLLVC (SEQ ID NO:42) or the peptide from SEQ ID NO:4 CEGLPPSTSSGQPPLQDMLCLGGVAVSLSHIRN (SEQ ID NO:43) or the

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peptide from SEQ ID NO:4 FMRYRSSSVLSHEEC (SEQ ID NO:44) . The peptides were synthesized using an Applied Biosystems Model 431A peptide synthesizer (Applied Biosystems, Inc., Foster City, CA) according to manufacturer's instructions. 5 The peptides were then conjugated to the carrier protein maleimide-activated keyhole limpet hemocyanin (KLH) through cysteine residues according to manufacturer's instructions. (Pierce, Rockford, IL). The rabbits were each given an initial intraperitoneal (IP) injection of 10 200 μ g of the conjugated peptide in Complete Freund's Adjuvant (Pierce, Rockford, IL) followed by booster IP injections of 100 μg conjugated peptide in Incomplete Freund's Adjuvant every three weeks. Seven to ten days after the administration of the third booster injection, 15 the animals were bled and the serum was collected. rabbits were then boosted and bled every three weeks.

The zalpha12 peptide-specific Rabbit seras were characterized by an ELISA titer check using 1 μg/ml of the peptide used to make the antibody as an antibody target.

20 The 2 rabbit seras to the SEQ ID NO:41 peptide have titer to their specific peptide at a dilution of 1:5E5 (1:100000). The 2 rabbit seras to the SEQ ID NO:42 peptide had titer to their specific peptide at a dilution of 1:5E4 (1:10,000). The 2 rabbit seras to the SEQ ID NO:43 peptide had titer to their specific peptide at a dilution of 1:5E4. The 2 rabbit seras to the SEQ ID NO:44 peptide had titer to their specific peptide at a dilution of 1:5E5.

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From the foregoing, it will be appreciated that, although specific embodiments of the invention have been described herein for purposes of illustration, various modifications may be made without deviating from the spirit and scope of the invention. Accordingly, the invention is not limited except as by the appended claims.

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CLAIMS

WHAT IS CLAIMED IS:

- 1. An isolated polypeptide comprised of an amino acid sequence selected from the group consisting of SEQ ID NOs.2, 4, 6, 8, 9, 10, 18, 19, 20, 25, 26, 27, 28, 29, 30, 35, 36, 37, 38, 39, 40, 41, 42, 43 and 44 or a polypeptide which is at least 80% identical to said polypeptide.
- 2. An isolated polypeptide comprised of an amino acid sequence selected from the group consisting of SEQ ID NOs.2, 4, 6, 8, 9, 10 or a polypeptide which is at least 70% identical to said polypeptide.
- 3. An isolated polynucleotide which encodes a polypeptide, wherein said polypeptide is comprised of an amino acid sequence selected from the group consisting of SEQ ID NOs. 2, 4, 6, 8, 9, 10, 18, 19, 20, 25, 26, 27, 28, 29, 30, 35, 36, 37, 38, 39 and 40 or a polypeptide which is at least 80% identical to said polypeptide.
- 4. An isolated polynucleotide which encodes a polypeptide, wherein said polypeptide is comprised of an amino acid sequence selected from the group consisting of SEQ ID NOs. 2, 4, 6, 8, 9 and 10 or a polypeptide which is at least 70% identical to said polypeptide.
- 5. An antibody which specifically binds to a polypeptide, wherein said polypeptide is comprised of an amino acid sequence selected from the group consisting of SEQ ID NOs. 2, 4, 6, 8, 9, 10, 18, 19, 20, 25, 26, 27, 28, 29, 30, 35, 36, 37, 38, 39 and 40.
- 6. An antibody of claim 5 which specifically binds to a polypeptide, wherein said polypeptide is comprised of an

amino acid sequence selected from the group consisting of SEQ ID NOs. 2, 4, 6, 8, 9, and 10.

- 7. An anti-idiotypic antibody which specifically binds to an antibody which specifically binds to a polypeptide, wherein said polypeptide is comprised of an amino acid sequence selected from the group consisting of SEQ ID NOs. 2, 4, 6, 8, 9, 10, 18, 19, 20, 25, 26, 27, 28, 29, 30, 35, 36, 37, 38, 39 and 40.
- 8. An anti-idiotypic antibody of claim 7 wherein the polypeptide is comprised of an amino acid sequence selected from the group consisting of SEQ ID NOs. 2, 4, 6, 8, 9 and 10.
- 9. The use of an antagonist to a polypeptide for treating inflammation wherein the polypeptide is comprised of an amino acid sequence selected from the group consisting of SEQ ID NOs. 2, 4, 6, 8, 9 and 10.
- 10. The use of claim 9 wherein the antagonist is an antibody which binds to a polypeptide comprised of an amino acid sequence selected from the group consisting of SEQ ID NOs. 2, 4, 6, 8, 9 and 10.
- 11. The use of an antagonist to a polypeptide for manufacturing a medicament for treating inflammation wherein the polypeptide is comprised of an amino acid sequence selected from the group consisting of SEQ ID NOs. 2, 4, 6, 8, 9 and 10.
- 12. The use of claim 11 wherein the antagonist is an antibody which binds to a polypeptide comprised of an amino acid sequence selected from the group consisting of SEQ ID NOs. 2, 4, 6, 8, 9 and 10.

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1117

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		PC1/US	5 00/01015
A CLASS	ification of subject matter C07K14/52 C12N15/19 C07K16/	/24 A61K38/19 A	A61P43/00
· · · · · · · ·	o International Patent Classification (IPC) or to both national classification	ication and IPC	
	SEARCHED		
IPC 7	ocumentation searched (classification system followed by classification sy	tion symbols)	
Documenta	tion searched other than minimum documentation to the extent that	such documents are included in the fi	leide searched
Flectmole d	lata base consulted during the International search (name of data b	•	
	and some constant daming the threshearth is seen on (harrier of case of	esse and, where practical, search term	as used)
C. DOCUM	ENTS CONSIDERED TO BE RELEVANT		
Category •	Citation of document, with indication, where appropriate, of the re	elevent passages	Relevant to claim No.
Х	DATABASE EMBL NUCLEOTIDE AND PRO SEQUENCES,14 May 1997 (1997-05-1 XP002138914	TEIN 4),	3,4
	HINXTON, GB AC = AA419437. zv01g08.r1 NCI_CG Homo sapiens cDNA clone IMAGE:74	AP_GCB1 6270 5'.	
_	mRNA sequence. EST.	·	
A	WO 95 13393 A (CELL THERAPEUTICS 18 May 1995 (1995-05-18) the whole document	INC)	1
A	WO 93 21308 A (UNIV HOSPITAL) 28 October 1993 (1993-10-28) the whole document		1
		_	
		-/	•
	ner documents are listed in the continuation of box C.	Patent family members are	fated in annex.
" Special cat	tegories of cited documents :	"I later document published after th	e international filing date
"E" earlier d	int defining the general state of the art which is not ared to be of particular relevance locument but published on or after the international	or priority date and not in confile cited to understand the principle invention "X" document of particular relevance	x with the application but or theory underlying the
filing da "L" documen	nt which may throw doubts on priority claim(a) or	cannot be considered novel or o	cannot be considered to
citation	is cust to establish the publication date of another i or other special reason (as specified)	"Y" document of particular relevance	the claimed invention
"O" docume	int referring to an oral disclosure, use, exhibition or nears	cannot be considered to involve document is combined with one ments, such combination being	or more other such docu-
sector th	nt published prior to the international filing date but an the priority date claimed	in the art. "&" document member of the same p	· ·
	actual completion of the International search	Date of mailing of the Internation	hal search report
	9 May 2000	14/06/2000	
Name and m	asking address of the ISA European Patent Office, P.B. 5618 Patentiaan 2 NL - 2280 HV Rijswijk	Authorized officer	
	Tel. (+31-70) 340-2040, Tx. 31 651 epo ni, Fex: (+31-70) 340-3016	Mateo Rosell,	A.M.

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Category *	ation) DOCUMENTS CONSIDERED TO BE RELEVANT Citation of document, with indication, where appropriate, of the relevant passages	To .	
	от по поменти развадев	Hei	evant to claim No.
Ρ,Χ	WO 99 29720 A (ZYMOGENETICS INC) 17 June 1999 (1999-06-17) the whole document		1,6-12
Ρ,Χ	DATABASE EMBL NUCLEOTIDE AND PROTEIN SEQUENCES,1 May 1999 (1999-05-01), XP002138913 HINXTON, GB AC = 095505. TrEMBLrel. DJ821D11.1 (PUTATIVE PROTEIN) (FRAGMENT). From nt 1-51. abstract		1-4
Ρ,Χ	DATABASE EMBL NUCLEOTIDE AND PROTEIN SEQUENCES,30 November 1999 (1999-11-30), XP002138915 HINXTON, GB AC = AW195720. xn85b08.x1 Soares_NFL_T_GBC_S1 Homo sapiens cDNA clone IMAGE:2701239 3', mRNA sequence. EST. abstract		3,4
Р,Х	DATABASE EMBL NUCLEOTIDE AND PROTEIN SEQUENCES,2 August 1999 (1999-08-02), XP002138916 HINXTON, GB AC = AI928166. wp11h06.x1 NCI_CGAP_Kid12 Homo sapiens cDNA clone IMAGE:2464571 3', mRNA sequence. EST. abstract		3,4

International application No.

PCT/US 00/01015

Box I Observations where certain claims were found unsearchable (Continuation of Item 1 of first sheet)
This international Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:
Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely:
2. Claims Nos.:
because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful international Search can be carried out, specifically:
3. Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).
Box il Observations where unity of invention is tacking (Continuation of item 2 of first sheet)
This international Searching Authority found multiple inventions in this international application, as follows:
į
As all required additional search fees were timely paid by the applicant, this international Search Report covers all
searchable dains.
2. As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:
The state of the s
 No required additional search fees were timely paid by the applicant. Consequently, this international Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:
and the state of t
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Remark on Protest The additional search fees were accompanied by the applicant's protest.
No protest accompanied the payment of additional search fees.
the product accompanies use payment of assessed accompanies

International Application No. PCT/US 00 \(D1015 \)

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

Continuation of Box I.2

Claims Nos.: 9,11

Present claims 9 and 11 relating to the use of an antagonist to the polypeptides comprised in the SEQ.ID.Nos. 2,4,6,8,9 and 10, could not be searched as its subject-matter was insuficiently disclosed.

The applicant's attention is drawn to the fact that claims, or parts of claims, relating to inventions in respect of which no international search report has been established need not be the subject of an international preliminary examination (Rule 66.1(e) PCT). The applicant is advised that the EPO policy when acting as an International Preliminary Examining Authority is normally not to carry out a preliminary examination on matter which has not been searched. This is the case irrespective of whether or not the claims are amended following receipt of the search report or during any Chapter II procedure.

information on patent family members

Intern 1at Application No PCT/US 00/01015

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